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Original article

X-ray tomography as a tool for detailed anatomical analysis

Jan VAN DEN BULCKE[1]*, Matthieu BOONE[2], Joris VAN ACKER[1], Marc STEVENS[1], Luc VAN HOOREBEKE[2]

[1] Laboratory of Wood Technology, Faculty of Bioscience Engineering, Ghent University, Coupure Links 653, 9000 Ghent, Belgium
[2] Department for Subatomic and Radiation Physics, Faculty of Sciences, Ghent University, Proeftuinstraat 86, 9000 Gent, Belgium

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Abstract

• Wood identification, anatomical examination and retrieval of quantitative information are important aspects of many research disciplines. Conventional light microscopy with a camera and (semi)-automatic image analysis software is an often used methodology for these purposes. More advanced techniques such as fluorescence, scanning electron, transmission electron, confocal laser scanning and atomic force microscopy are also part of the toolset answering to the need for detailed imaging.

• Fast, non-destructive visualization in three dimensions with high resolution combined with a broad field of view is sought-after, especially in combination with flexible software.

• A highly advanced supplement to the existing techniques, namely X-ray sub-micron tomography, meets these requirements. It enables the researcher to visualize the material with a voxel size approaching < 1 μm for small samples (< 1 mm). Furthermore, with tailor-made processing software quantitative data about the wood in two and three dimensions can be obtained. Examples of visualization and analysis of four wood species are given in this paper, focusing on the opportunities of tomography at micron and sub-micron resolution.

• X-ray computed tomography offers many possibilities for material research in general and wood science in specific, as a qualitative as well as a quantitative technique.

Mots-clés :
RX CT /
bois /
anatomie /
traitement d'images

Résumé – La micro-tomographie RX, un outil pour une analyse anatomique fine du bois.

• L'identification du bois, l'observation anatomique et l'obtention d'informations quantitatives sont des aspects importants dans différentes disciplines scientifiques. La microscopie optique conventionnelle couplée à l'acquisition et au traitement semi automatique des images est souvent utilisée pour atteindre ces objectifs. Des techniques plus récentes comme la fluorescence, la microscopie électronique par balayage ou par transmission, la microscopie confocale ou encore à force atomique constituent une panoplie d'outils répondant à ces besoins d'imagerie fine.

• Il y a une forte demande d'outils non destructifs de visualisation 3D à haute résolution combinés à un large champ de vision et surtout avec des logiciels flexibles.

• En complément avancé à ces techniques la tomographie RX submicrométrique remplit ces conditions. Elle permet au chercheur de visualiser le matériau avec une taille de voxel inférieure au micron pour de petits échantillons dont la taille est inférieure au mm. En outre, à l'aide de logiciels de traitement adaptés, des données quantitatives peuvent être obtenues pour le bois en deux et trois dimensions. Dans ce papier on présente des exemples de visualisation et d'analyse pour quatre essences en focalisant sur les possibilités de tomographie aux échelles micrométrique et submicrométrique.

• La tomographie RX offre de nombreuses possibilités pour la recherche en science des matériaux en général et en sciences du bois en particulier, que ce soit pour les approches qualitatives ou pour les approches quantitatives.

* Corresponding author: Jan.VandenBulcke@UGent.be

1. INTRODUCTION

Wood identification as well as quantification of anatomical features is important for many disciplines such as tree physiology (Fonti et al., 2007), wood technology (Makinen et al., 2008), archaeology (Philippe and Bamford, 2008), forensics (Coyle et al., 2001), etc. Identification relies on the macroscopic appearance and the characteristics revealed under a microscope. Mainly, examining axial, tangential and radial microtome sections is necessary for correct determination. Semi-automated image analysis of these sections leads to quantitative data such as porosity, fibre length, vessel diameter, cell wall thickness etc. The conventional approach consists of a microtome, a light microscope with a camera mounted on top and image analysis software. Modern techniques can assist in determination and characterization of wood species: SEM (Scanning Electron Microscopy), TEM (Transmission Electron Microscopy), AFM (Atomic Force Microscopy), CSLM (Confocal Scanning Laser Microscopy) etc. Yet most of the time, obtaining quantitative information is labour- and time intensive. In addition to aforementioned techniques, X-ray tomography is explored in this paper as a tool for detailed anatomical research. It is a technique used in several research disciplines such as medicine (Fu and Kuduvalli, 2008), soil science (Taina et al., 2008), hydrology (Wildenschild et al., 2002), entomology (Fuchs et al., 2004), plant physiology (Lee and Kim, 2008) and material science (Cnudde and Jacobs, 2004) to name only a few. Even in wood science, its possibilities are employed. In its two dimensional form, X-ray analysis is already used for densitometry (Knapic et al., 2007; Macchioni et al., 2007; Tomazello et al., 2008). X-ray computed tomography in three dimensions is utilized for the analysis of low-density fibreboard under compression (Badel et al., 2008), study of wood-plastic composites (Wang et al., 2007), detection of organosilicon compounds (De Vetter et al., 2006), microstructure analysis of spruce wood (Trtik et al., 2007) and quantitative wood anatomy (Steppe et al., 2004).

The purpose of this article is to illustrate the power of X-ray computed tomography as a tool for both descriptive and quantitative wood identification and anatomy to resolve details on three-dimensional reconstructions with near sub-micron scale without destruction or labour-intensive sample preparation. This non-destructiveness has the advantage to visualize the object's original structure, without cell damage or artefacts during sample preparation. What is more, the flexible set-up allows scanning of objects of diverse dimensions with a sufficient field of view resulting in large high detailed volumes. As such, the sample can be examined in all possible directions, making fast evaluation possible. Parallax effects as explained by Park and Telewski (1993) cause no problems and any manipulation of the virtual object is possible. The technique is illustrated for four wood species using several self-explanatory images and calculations of cell wall thickness and cell lumen size on 2D slices with standard MATLAB® algorithms. To highlight the possibility of 3D quantitative wood anatomy, a subvolume of a data stack is processed. Special software (Morpho+; Vlassenbroeck et al., 2007) for handling of large datasets is demonstrated as well.

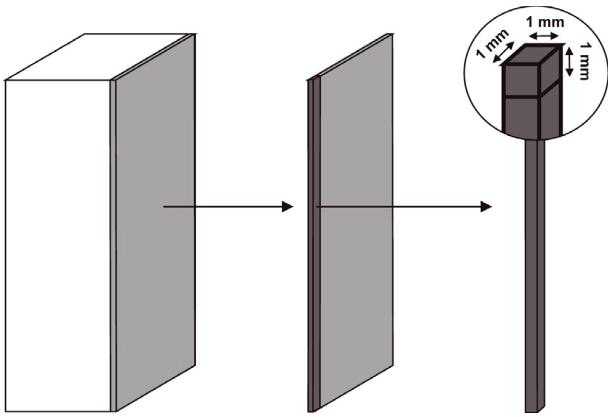

Figure 1. Sample preparation for X-ray scanning.

2. MATERIALS AND METHODS

The four wood species used for X-ray analysis are Scots pine (*Pinus silvestris* L. – earlywood and latewood), beech (*Fagus sylvatica* L.), movingui (*Disthemonanthus benthamianus* Baill.) and afzelia (*Afzelia bipindensis* Harms). These species represent hard- and softwood as well as temperate and tropical wood species. Pine sapwood and beech are often used in European standards whereas movingui and afzelia are durable tropical species on the market (e.g. in Belgium). Five samples, two for pine (early- and latewood) and one per other wood species, were prepared by slicing a thin wood section of a larger block and subdividing it with a microtome or scalpel in needle-shaped specimens (Fig. 1).

The tip of this needle-shaped wood sample, measuring approximately one mm^3 was scanned using the X-ray equipment built at the Centre for X-ray Tomography at Ghent University (UGCT, http://www.ugct.ugent.be). This is a state-of-the-art scanner (Masschaele et al., 2007), highly flexible, with in-house developed software for scanner control, sample reconstruction, analysis and visualization. The X-ray source, a nano-focus tube, can reach a focal spot size down to one μm. All samples were scanned at an average voltage of 50 kV and a current of 40 μA with a total scan time of approximately 2 h. A rotation step size of 0.36° was used. Reconstruction took 20 min with Octopus, a server/client tomography reconstruction package for parallel and cone beam geometry (Vlassenbroeck et al., 2007). With the described set-up submicron resolution can be reached, resulting in scans with voxels sizing approximately $0.7 \times 0.7 \times 0.7$ μm. The small voxel size gave a clear view on anatomical features. Subvolumes of these reconstructed slices were further manipulated with MATLAB® and Morpho+ (Vlassenbroeck et al., 2007). First, the slices were pre-processed aiming at noise removal and image enhancement. This included histogram equalization to transform the values of the greyscale images such that contrast was improved. Subsequently the images were binarized using the topological derivative of Larrabide (2008). Finally, slices were despeckled by removal of small isolated pixel islands. For the pine early- and latewood specimen, better results were obtained when images were denoised using the non-linear diffusion technique as outlined by D'Almeida (non-linear diffusion toolbox by Frederico D'Almeida). Subsequently, once denoised images were available, wood parameters such as cell wall thickness and cell lumen size could be calculated on 2D slices and labelling of a subvolume of pine (Van den Bulcke et al., 2008) could be performed in three dimensions. Cell lumen sizes were determined via

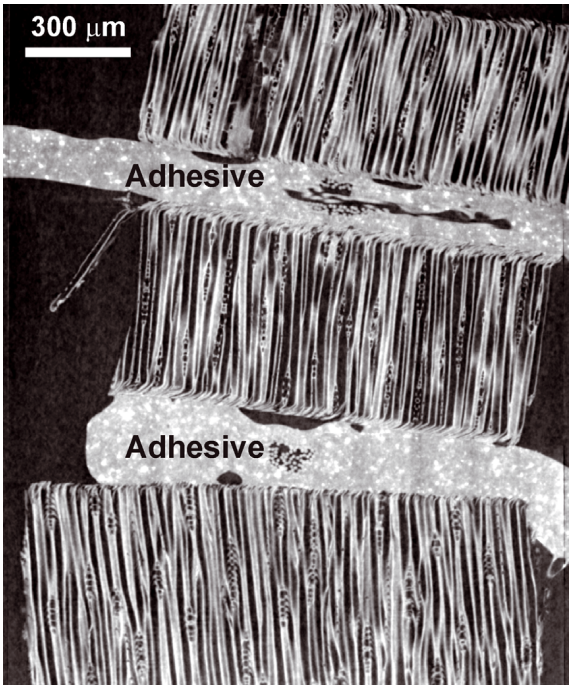

Figure 2. Cross-sectional view through a stack of three pine latewood samples.

marker-based segmentation. This procedure starts with determination of the local maxima of the distance transform of the image, as such representing the centres of the cell lumens. Application of the watershed algorithm with these local maxima as markers correctly separated cells formerly connected though open pits. In fact, this is an automatic version of the technique described in Reme and Helle (2002) for pit removal. Manual editing was necessary to remove ray cells from analysis and to undo incorrect segmentation, but analysis is quite fast, naturally depending on the quality of segmentation. Calculation of cell wall thickness was accomplished by using the distance transform of the skeletonised image. The cell wall thickness (CW) then equals (1):

$$CW = \frac{2 \times r \times \sum_{i=1}^{m} \sum_{j=1}^{p} (skel(i,j) \times dist(i,j))}{\sum_{i=1}^{m} \sum_{j=1}^{p} skel(i,j)} \quad (1)$$

with r = resolution (µm)
$skel$ = skeletonised cell walls
$dist$ = distance transformed cell walls
i, j = row and column indices
m, p = row and column size of matrix.

In addition, to exemplify the practical use of X-ray computed tomography in wood research, six pine latewood volumes, sampled from pith to bark, were scanned. A modified bronnikov algorithm (Boone et al., 2009) was employed for phase-contrast imaging. Figure 2 shows a slice through one of the two scanned stacks consisting of three samples separated by an adhesive tape. The voxel size was 1.68 µm and samples measured approximately 1.6 × 1.8 × 1.1 mm.

For correct analysis, rotation of the samples was obligatory. Once rotated, preprocessing included smoothing, noise removal and automated greyscale thresholding. Analysis of several 2D sections per sample resulted in mean values for cell perimeter and cell wall thickness and a profile in function of age.

All images were rendered with VGStudio MAX®, MATLAB®, Octopus 3D Viewer and Drishti (Limaye, 2006).

3. RESULTS AND DISCUSSION

For each wood species, several images will illustrate the anatomy. A 3D reconstruction gives an overview of the scan, cross-sectional views in axial, radial and tangential direction are given similar to conventional cross-sectional views by Wagenführ and Schreiber (1989), IAWA list of microscopic features for hardwood identification (IAWA Committee et al., 1989), Schweingruber (1990), IAWA list of microscopic features for softwood identification (IAWA Committee et al., 2004) and Wagenführ (2007). Characteristics of the wood are calculated on 2D sections and a 3D subvolume is used for 3D analysis. Finally, the brief study of six pine latewood samples further exemplifies the practical use of X-ray tomography in wood research.

3.1. Microscopic features

Below follows an overview of the characteristics of the wood species under study.

Pinus silvestris L.

Scots pine, member of the Pinaceae family, is characterized by its homogeneous anatomy with tracheids as the main structural element and with a minor share of resin channels. Early- (EW) and latewood (LW) are clearly different in tracheid wall thickness and lumen size. Pitting of radial walls of tracheids is predominantly uniseriate. Wood rays are heterocellular, composed of ray parenchyma and ray tracheids with dentate thickenings and small pits. Cross-field pitting is fenestriform, on average one pit per cross-field. These microscopic features can be visualized on 3D reconstructions of pine given in Figure 3 and several views through the volume.

Fagus sylvatica L.

Beech, member of the Fagaceae family, is a temperate hardwood species with a typical diffuse porous structure. The vessels are scattered and only reach small diameters. Wood rays can be subdivided in large and small individuals. Fibres have a relatively small lumen and a moderate thick wall. Vessel ray pitting is horizontal and have much reduced borders (predominantly scalariform). The compilation of images in Figure 4 illustrates several of these characteristics.

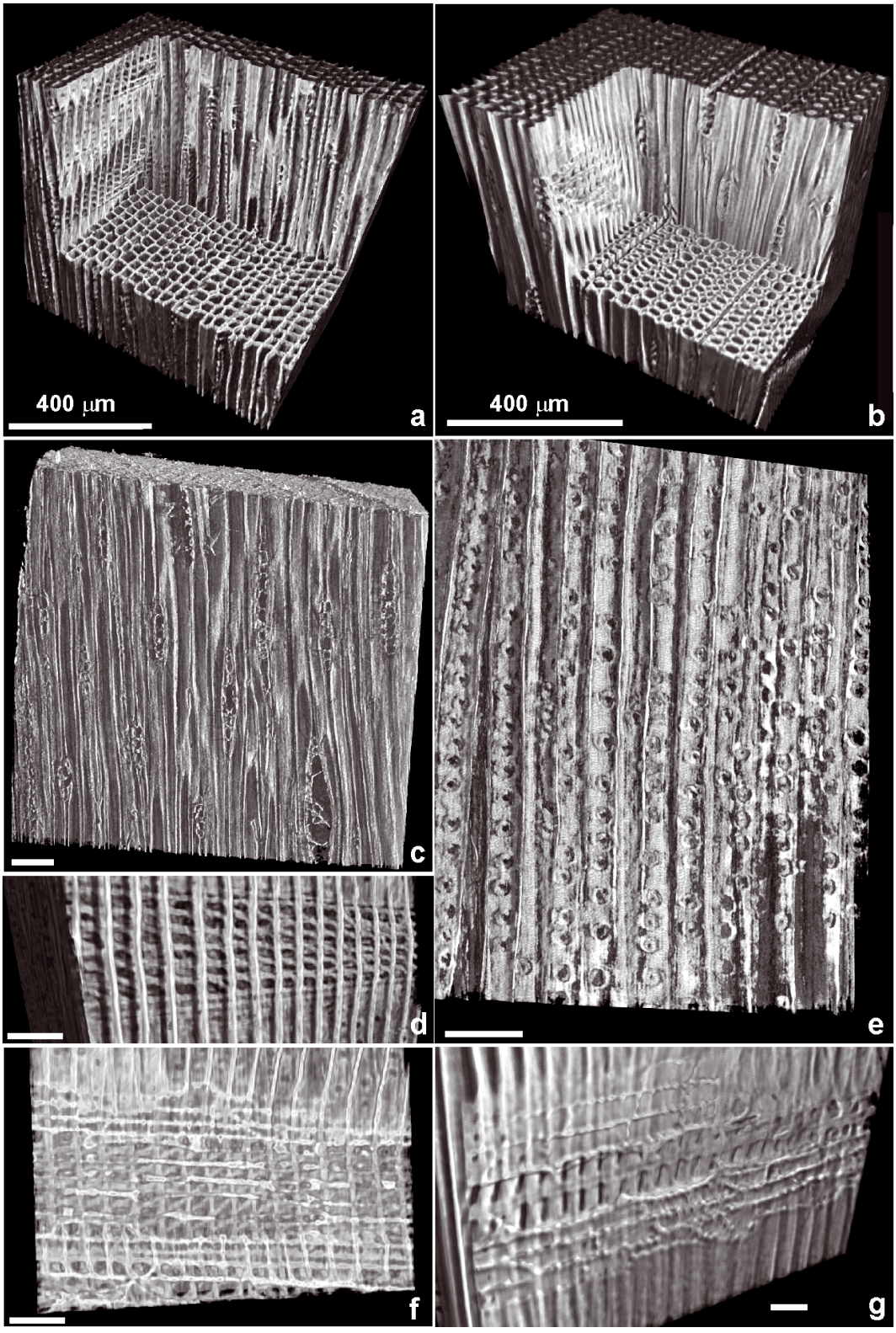

Figure 3. Overview with cut out giving a clear view on the internal anatomy of *Pinus silvestris* L. (a) earlywood and (b) latewood; (c) rays in latewood with limited height; (d) fenestriform cross-field pitting; (e) uniseriate pitting of cell walls of tracheids; (f+g) dentation of cell walls adjacent to ray parenchyma. White bar = 100 µm.

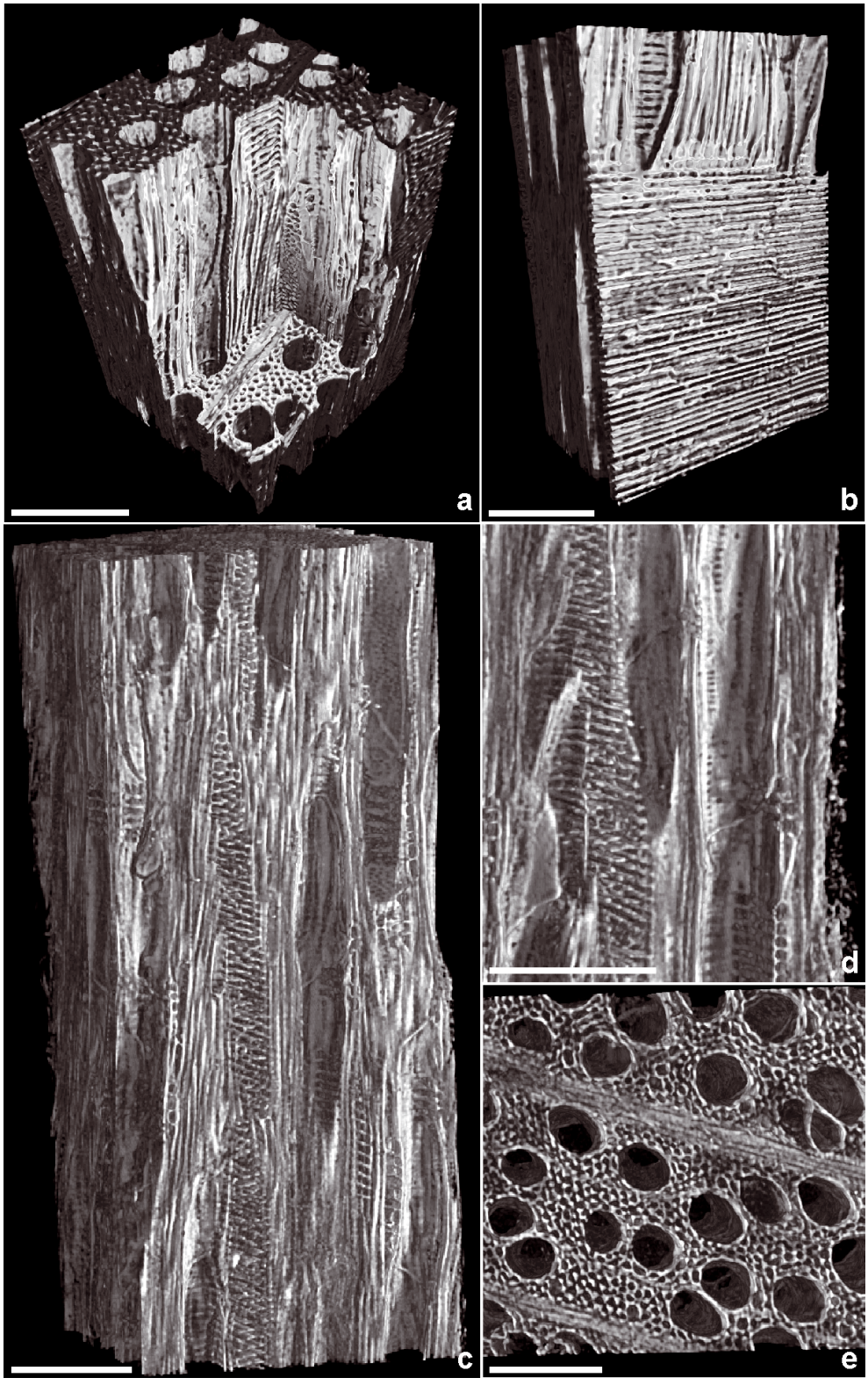

Figure 4. Overview with cut out (a) giving a clear view on the internal anatomy of *Fagus sylvatica* L.; (b) rays and scalariform vessel-ray pitting; (c) large rays with several vessel cross-sections; (d) perforations of the vessel wall and view on the ray frame; (e) top view. White bar = 200 μm.

Disthemonanthus benthamianus Baill.

Movingui, member of the Leguminosae-Caesalpinioideae family, is a diffuse-porous tropical hardwood, with little but large vessels and fibres. Parenchyma is apotracheal-terminal or paratracheal-aliform. Heterogeneous wood rays are small and not very high. The presence of silicon in heartwood is not unusual. Figure 5 illustrates the different anatomical aspects of movingui.

Afzelia bipindensis Harms

Afzelia, also a member of the Leguminosae-Caesalpinioideae family, is a diffuse-porous tropical hardwood, with little but large vessels and thick fibres. Parenchyma is paratracheal aliform. Wood rays are small and not very high. Vessel perforation is simple. Crystals are abundantly present. The rendered volumes in Figure 6 exemplify the wood anatomy.

3.2. Quantitative analysis

Image processing of large anisotropic volumes is a difficult task. Yet, wood consists of a 3D structure that can be described effectively with 2D sections. Therefore, for the determination of parameters such as cell (lumen) size and cell wall thickness, a 2D section is sufficient. The procedure is very straightforward once a noise-free image is obtained. Pre-processing included selection of the tissue of interest, histogram equalization, standard noise filtering and image binarization. As an example, Figure 7 illustrates the difference between the original images with noise, the noise-free (MATLAB®) and segmented (Morpho+) images for the four wood species. The segmented image can be used for measuring cell properties; in fact, the segmented images in Figure 7c are colour-coded according to their lumen size.

Via marker-based watershed image segmentation cell lumen size was determined. Cell lumen size frequencies are given in the line plots in Figure 8 for the four wood species. The vessel sizes of afzelia and movingui are not displayed because of their large size and low frequency. It should be mentioned that, considering the natural variability of wood anatomy, these results are not representative of the four species.

The difference in lumen size between early- and latewood tracheids is clear. Larger structures such as rays were labelled as well and manually removed from the analysis, but could be filtered (semi-)automatically using shape descriptors, e.g. by their elongated form. Dimensions can be used to split up vessels and fibres. It is clear that by proper demarcation of the different zones, the different tissue sizes can be determined. Once such a segmented and labelled dataset is available, calculations of a whole set of properties is straightforward. Cell wall thickness is calculated from these segmented images by skeletonization (Fig. 9). Results for pine are in agreement with the data in literature (Reme and Helle, 2002) while for the other species the reader is referred to Wagenführ and Schreiber (1989) and Wagenführ (2007).

As an example of 3D analysis performed in MATLAB®, the pine earlywood data stack is used. Optimal preprocessing of the stack of the original 2D sections included histogram equalization, noise removal by nonlinear image diffusion and image binarization using the topological derivative.

Three-dimensional reconstruction of a reduced region of interest based on watershed segmentation of the noise-free volume gives a view on the labelling of the different colour-shuffled tracheid lumens. The original binarized cell wall is visible as the greyish substance in-between in Figure 10a. This labelled volume can serve as the basis for calculation of length and shape of single elements. The limits of resolution permit to visualize individual pits as illustrated by Figures 10b and 10c. If 3D cell characteristics are desired, subtracting the skeletonised cell walls from the cell wall volume leads to the separated structures for volumetric analysis. However, whereas 2D analysis is fast, easy to correct and accurate 3D analysis of large volumes is much more difficult. Especially for hardwood species such analysis will pose problems and should be performed on isolated anatomical regions.

3.3. Analysis from pith to bark

A pith to bark analysis was performed for six pine latewood samples. Figure 11 illustrates the rendered volumes and the results of cell wall thickness and total cell perimeter measurements. These data are roughly in agreement with the data presented by Reme and Helle (2002). Obviously, juvenile wood has thinner walls and smaller cells in contrast with mature wood. It should be stressed that these data are not an in-depth study of the changing latewood characteristics of Scots pine yet a proof of concept of the use of the X-ray computed modality presented in this paper.

4. CONCLUSIONS

X-ray sub-micron tomography has been shown here to be a powerful image acquisition technique for wood research. The level of detail suffices for descriptive and quantitative anatomical analysis as exemplified in this paper. Very small samples can be scanned at very high resolution, making it appropriate for forensics and analysis of cultural heritage. Its non-destructiveness is an advantage when dealing with valuable material compared to classical methods, entailing the absence of preparation artefacts as well. Furthermore, volume mosaicing will enable the reconstruction of larger samples at a high level of detail, only limited by data handling and storage. In addition, a factor not to be neglected is the educational value of 3D images, which could complete the online databases such as InsideWood (http://insidewood.lib.ncsu.edu/search) and wood anatomy of Central European species (http://www.wsl.ch/land/products/dendro). At last, once the substrate

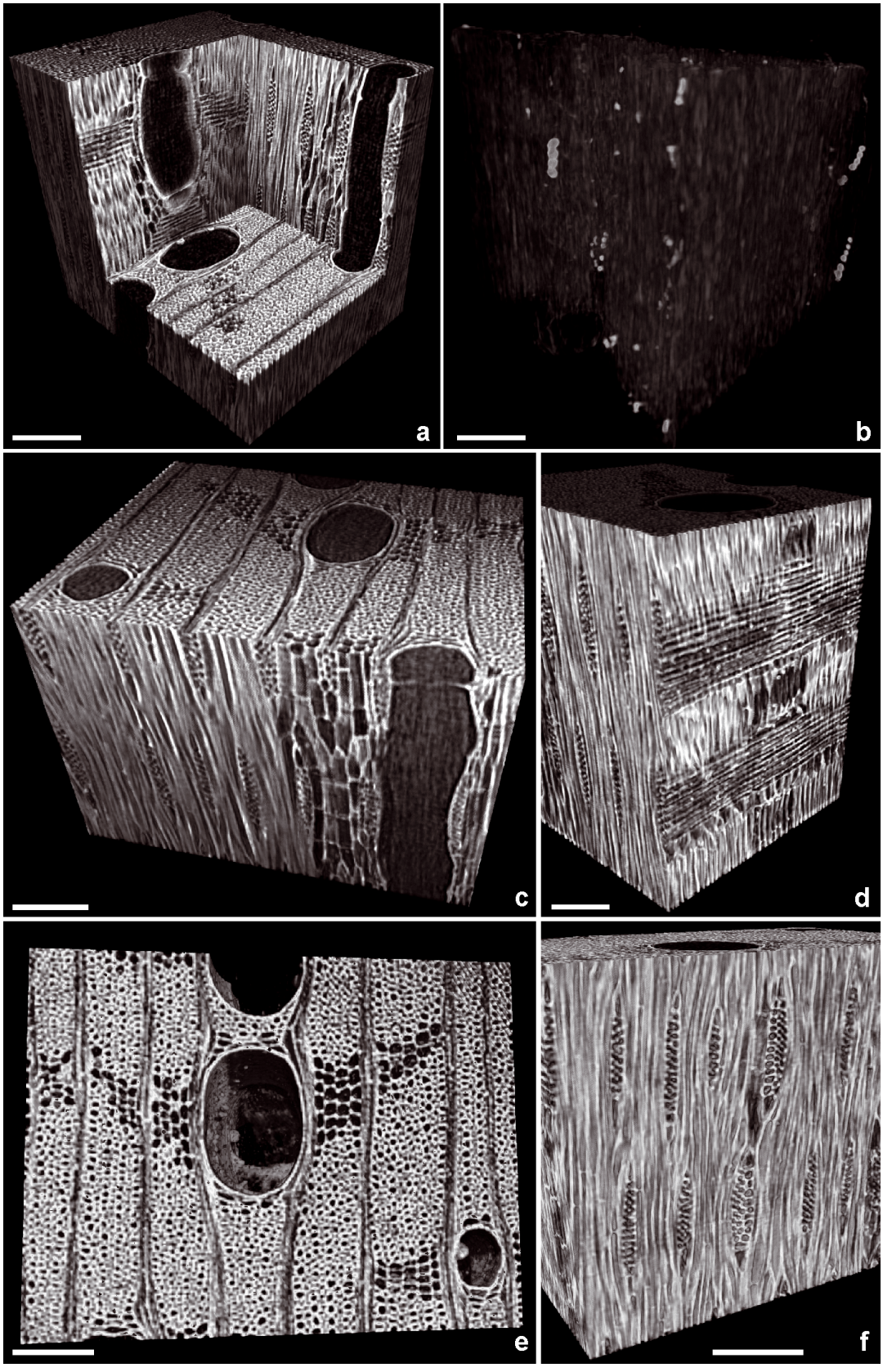

Figure 5. Overview with cut out (a) giving a clear view on the internal anatomy of *Disthemonanthus benthamianus* Baill.; (b) prismatic crystals; (c) parenchyma, rays and slice through vessel; (d) longitudinal slice through rays; (e) top view; (f) tangential view on rays. White bar = 200 μm.

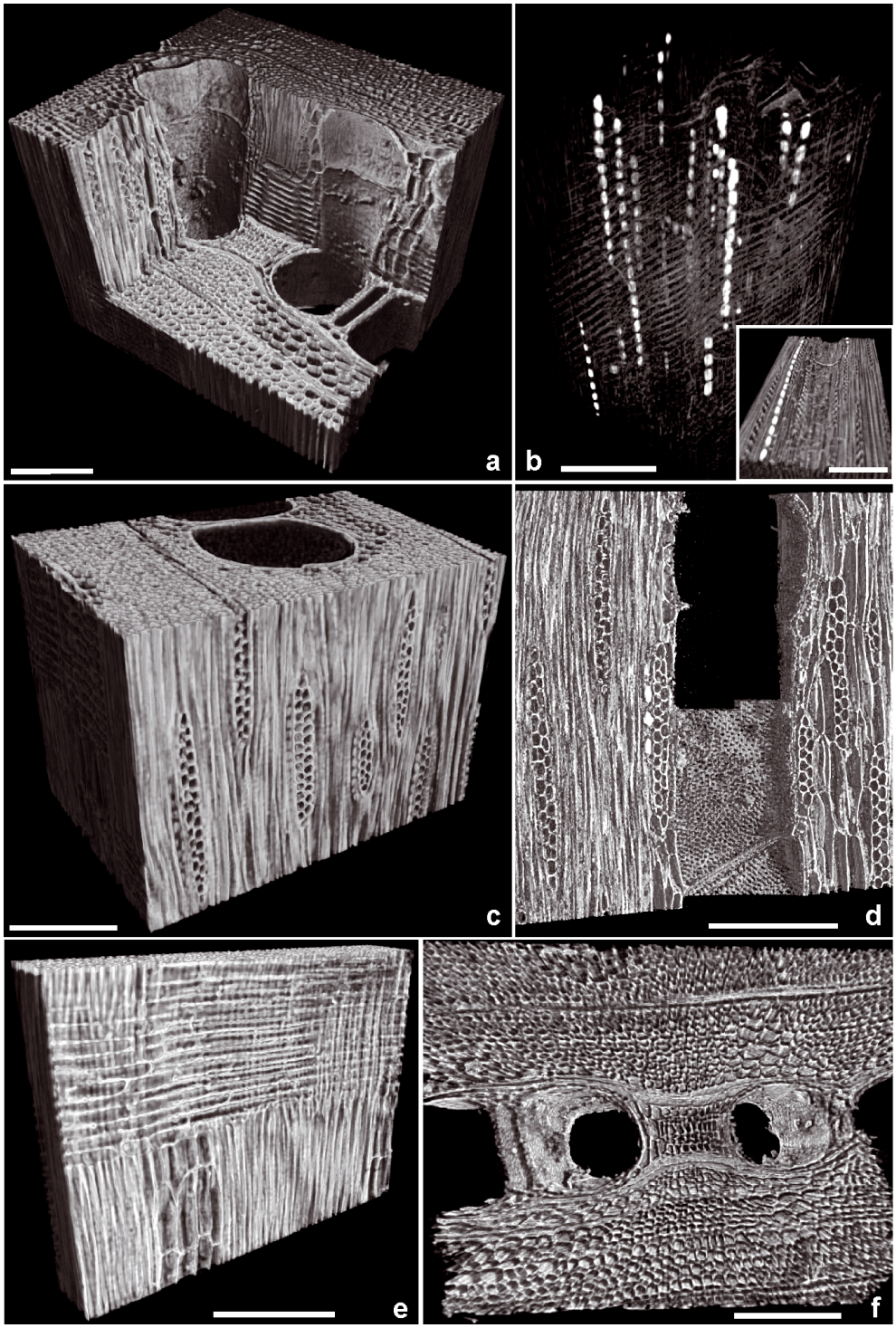

Figure 6. Overview with cut out (a) giving a clear view on the internal anatomy of *Afzelia bipindensis* Harms; (b) prismatic crystals; (c) rays; (d) simple vessel perforation; (e) view through ray and parenchyma; (f) top view. White bar = 200 μm.

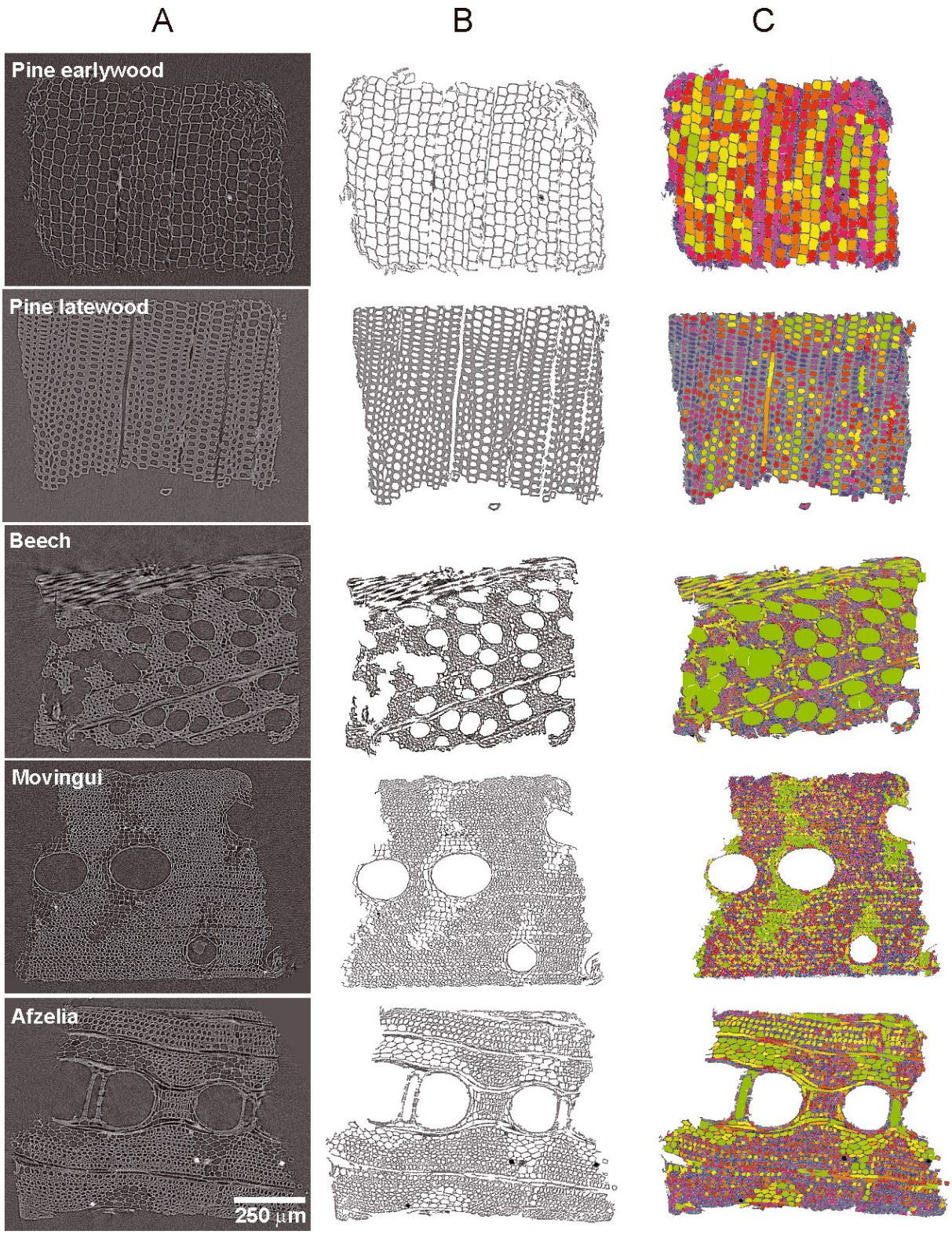

Figure 7. Original (A), noise-free (B) and segmented (C) images.

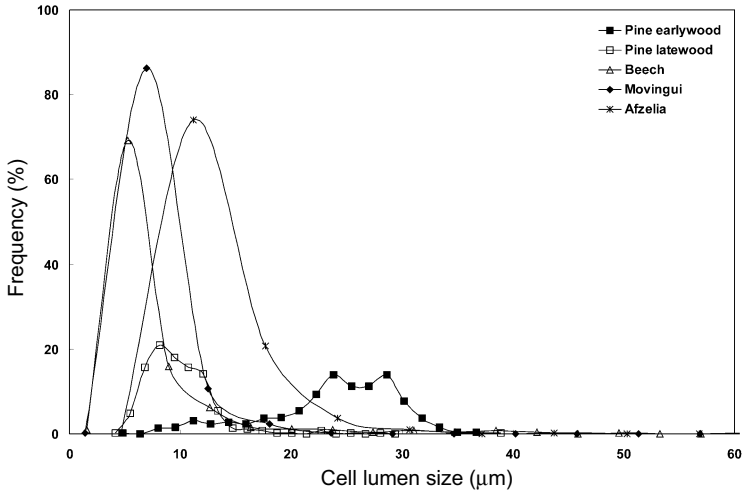

Figure 8. Frequencies of the equivalent diameter of the cell lumen sizes for four wood species.

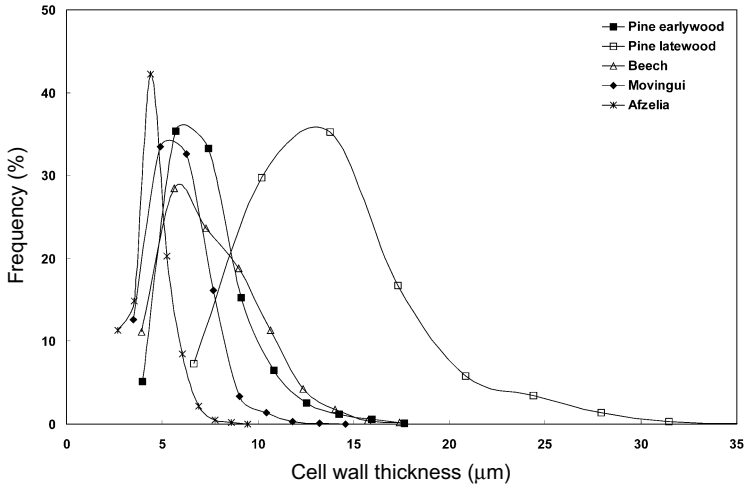

Figure 9. Frequencies of cell wall thicknesses for four wood species.

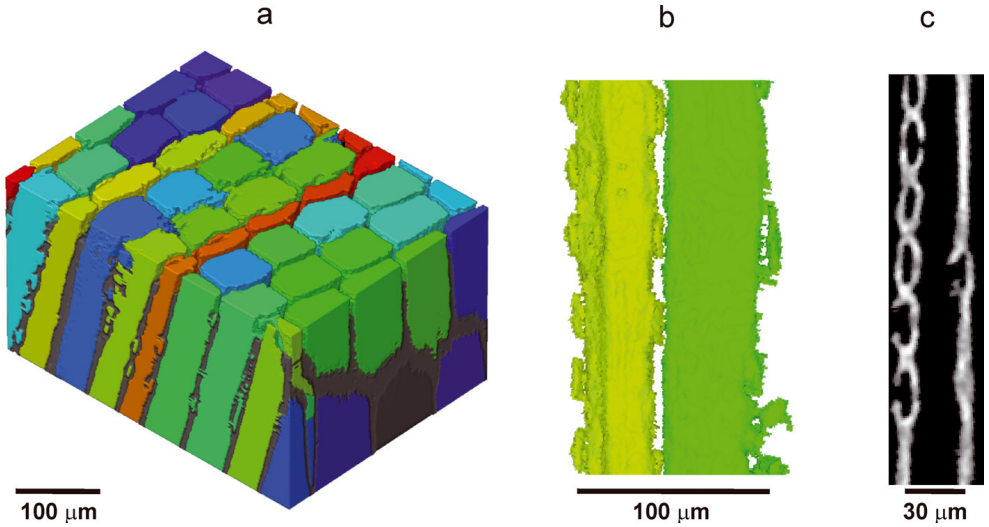

Figure 10. Labelled cell lumens of the noise-free pine earlywood sample with greyish cell wall (a), detailed rendering of pitting between two cells (b) and a 2D slice as an example of the limits of resolution for pit visualization.

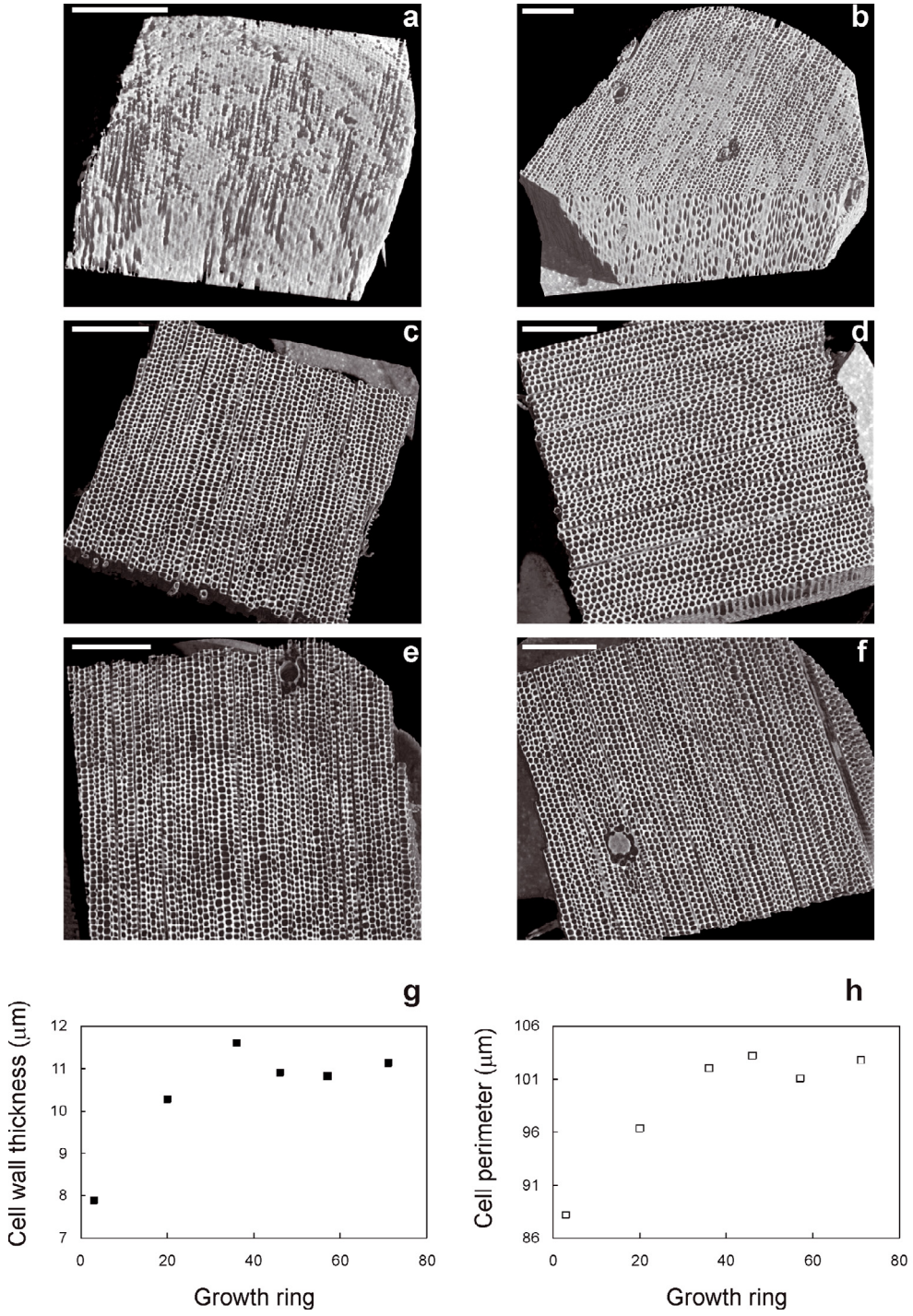

Figure 11. Six reconstructed pine latewood volumes sampled from pith to bark (a-f) and their cell wall thickness (g) and cell perimeter (h) in function of the growth ring they were sampled from. White bar = 400 μm.

is virtualized, all kinds of manipulation are feasible. The X-ray sub-micron tomography equipment presented in this paper will be a valuable tool in material and life sciences in general.

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REFERENCES

Badel E., Delisee C., and Lux J., 2008. 3D structural characterisation, deformation measurements and assessment of low-density wood fibreboard under compression: the use of X-ray microtomography. Compos. Sci. Technol. 68: 1654–1663.

Boone M., De Witte Y., Dierick M., Van den Bulcke J., Vlassenbroeck J., and Van Hoorebeke L. (2009). Practical use of the Modified Bronnikov algorithm in micro-CT. Nucl. Instrum. Methods Phys. Res. Sect. B: Beam Interact. Mater. At. 267: 1182–1186.

Cnudde V. and Jacobs P.J.S., 2004. Monitoring of weathering and conservation of building materials through non-destructive X-ray computed microtomography. Environ. Geol. 46: 477–485.

Coyle H.M., Ladd C., Palmbach T., and Lee H.C., 2001. The green revolution: Botanical contributions to forensics and drug enforcement. Croat. Med. J. 42: 340–345.

De Vetter L., Cnudde V., Masschaele B., Jacobs P.J.S., and Van Acker J., 2006. Detection and distribution analysis of organosilicon compounds in wood by means of SEM-EDX and micro-CT. Mater. Charact. 56: 39–48.

Fonti P., Solomonoff N., and Garcia-Gonzalez I., 2007. Earlywood vessels of Castanea sativa record temperature before their formation. New Phytol. 173: 562–570.

Fu D.S. and Kuduvalli G., 2008. A fast, accurate, and automatic 2D-3D image registration for image-guided cranial radiosurgery. Med. Phys. 35: 2180–2194.

Fuchs A., Schreyer A., Feuerbach S., and Korb J., 2004. A new technique for termite monitoring using computer tomography and endoscopy. Int. J. Pest Manage. 50: 63–66.

IAWA Committee, Richter H.G., Grosser D., Heinz I., and Gasson P.E., 2004. IAWA list of microscopic features for softwood identification. IAWA J. 25: 1–70.

IAWA Committee, Wheeler E.A., Baas P., and Gasson P.E., 1989. IAWA list of microscopic features for hardwood identification. IAWA Bull. 10: 219–332.

Knapic S., Louzada J.L., Leal S., and Pereira H., 2007. Radial variation of wood density components and ring width in cork oak trees. Ann. For. Sci. 64: 211–218.

Larrabide I., Feijóo R.A., Novotny A.A., and Taroco E.A., 2008. Topological derivative: A tool for image processing. Comput. Struct. 86: 1386–1403.

Lee S.J. and Kim Y., 2008. In vivo visualization of the water-refilling process in xylem vessels using X-ray micro-imaging. Ann. Bot. 101: 595–602.

Limaye A., 2006. Drishti – Volume exploration and presentation tool. visualization conference, Baltimore, USA.

Macchioni N., Palanti S., and Rozenberg P., 2007. Measurements of fungal wood decay on Scots pine and beech by means of X-ray microdensitometry, Wood Sci. Technol. 41: 417–426.

Makinen H., Jyske T., and Saranaa P., 2008. Variation of tracheid length within annual rings of Scots pine and Norway spruce. Holzforschung 62: 123–128.

Masschaele B.C., Cnudde V., Dierick M., Jacobs P., Van Hoorebeke L., and Vlassenbroeck J., 2007. UGCT: new X-ray radiography and tomography facility, Nucl. Instrum. Methods Phys. Res. Sect. A: Accel. Spectrom. Detect. Assoc. Equip. 580: 266–269.

Park W.-K. and Telewski F.W., 1993. Measuring maximum latewood density by image analysis at the cellular level. Wood Fiber Sci. 25: 326–332.

Philippe M. and Bamford M.K., 2008. A key to morphogenera used for Mesozoic conifer-like woods. Rev. Palaeobot. Palynol. 148: 184–207.

Reme P.A. and Helle T., 2002. Assessment of transverse dimensions of wood tracheids using SEM and image analysis, Holz Roh- Werkst. 60: 277–282.

Schweingruber F.H., 1990. Anatomy of European woods, Paul Haupt Berne and Stuttgart Publishers, Stuttgart.

Steppe K., Cnudde V., Girard C., Lemeur R., Cnudde J.P., and Jacobs P., 2004. Use of X-ray computed microtomography for non-invasive determination of wood anatomical characteristics. J. Struct. Biol. 148: 11–21.

Taina I.A., Heck R.J., and Elliot T.R., 2008. Application of X-ray computed tomography to soil science: A literature review. Can. J. Soil Sci. 88: 1–20.

Tomazello M., Brazolin S., Chagas M.P., Oliveira J.T.S., Ballarin A.W., and Benjamin C.A., 2008. Application of X-ray technique in non-destructive evaluation of eucalypt wood, Maderas-Cienc. Tecnol. 10, 139–149.

Trtik P., Dual J., Keunecke D., Mannes D., Niemz P., Stahli P., Kaestner A., Groso A., and Stampanoni M., 2007. 3D imaging of microstructure of spruce wood. J. Struct. Biol: 159, 46–55.

Van den Bulcke J., Masschaele B., Dierick M., Van Acker J., Stevens M., and Van Hoorebeke L., 2008. Three-dimensional imaging and analysis of infested coated wood with X-ray submicron CT. Int. Biodeterior. Biodegrad. 61: 278–286.

Vlassenbroeck J., Dierick M., Masschaele B., Cnudde V., Van Hoorebeke L., and Jacobs P., 2007. Software tools for quantification of X-ray microtomography at the UGCT. Nucl. Instrum. Meth. Phys. Res. A 580: 442–445.

Wagenführ R., 2007. Holzatlas, Fachbuchverlag Leipzig, Leipzig.

Wagenführ R. and Schreiber C., 1989. Holzatlas, 3rd éd. VEB Fachbuchverlag Leipzig, Leipzig.

Wang Y., Muszynski L., and Simonsen J., 2007. Gold as an X-ray CT scanning contrast agent: Effect on the mechanical properties of wood plastic composites. Holzforschung 61: 723–730.

Wildenschild D., Hopmans J.W., Vaz C.M.P., Rivers M.L., Rikard D., and Christensen B.S.B., 2002. Using X-ray computed tomography in hydrology: systems, resolutions, and limitations. J. Hydrol. 267: 285–297.