

## Comparison between the structure and function of chloroplasts at different levels of willow canopy during a growing season

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### Introduction

Light climate has a strong impact on the ultrastructure of chloroplasts. There is plenty of evidence that the degree of grana stacking in chloroplasts of plants grown in high light is less than in plants grown in low light (*e.g.*, Lichtenthaler *et al.*, 1981), which is also the case for plants adapted to sunny or shady habitats (Boardman, 1977; Aro *et al.*, 1986). Very little is, however, known about the seasonal acclimation process of the photosynthetic apparatus in the canopy, where leaves that are initially exposed to full sunlight are transferred through half-shade into full shade. In conditions, under which water and nutrient availability are not limiting growth, the shaded leaves remain intact for most of the growing season. This suggests that the leaves retain a positive carbon balance by acclimating to the changing light climate. In this study, we quantified the seasonal changes in the chloroplast ultrastructure at several heights of a willow (*Salix cv. Aquatica gigantea*) canopy. We also determined how changes in chloroplast ultrastructure

fit with their function by measuring the rate of gas exchange under the prevailing environmental conditions in the canopy.

### Materials and Methods

The willow stand (established in 1980, 125 m<sup>2</sup> in area) was cut down before the growing season 1986 and measurements were made on leaves that emerged on new shoots successively throughout the growing season. The stand was fertilized with a commercial fertilizer (Putarhan Y-lannos 10-16-17) once a week during the growing season, so that it received a total of 150 kg of N/ha/season. The stand was watered regularly to assure that the plants were not water-stressed.

The samples for electron microscopic examination were taken from 3 replicate plots at 6 different dates from upto 5 different heights (Fig. 1A). The samples were treated as described by Vapaavuori (1986) and Aro *et al.* (1986). The grids were examined on a Jeol 100B electron microscope. Before prefixation of the samples for electron microscopy, the photosynthetic capacity of the leaves was measured at prevailing light and temperature conditions by means of a CO<sub>2</sub> porometer (ADC LCA-2, the Analytical Development Co. Ltd., U.K.). The chloroplast

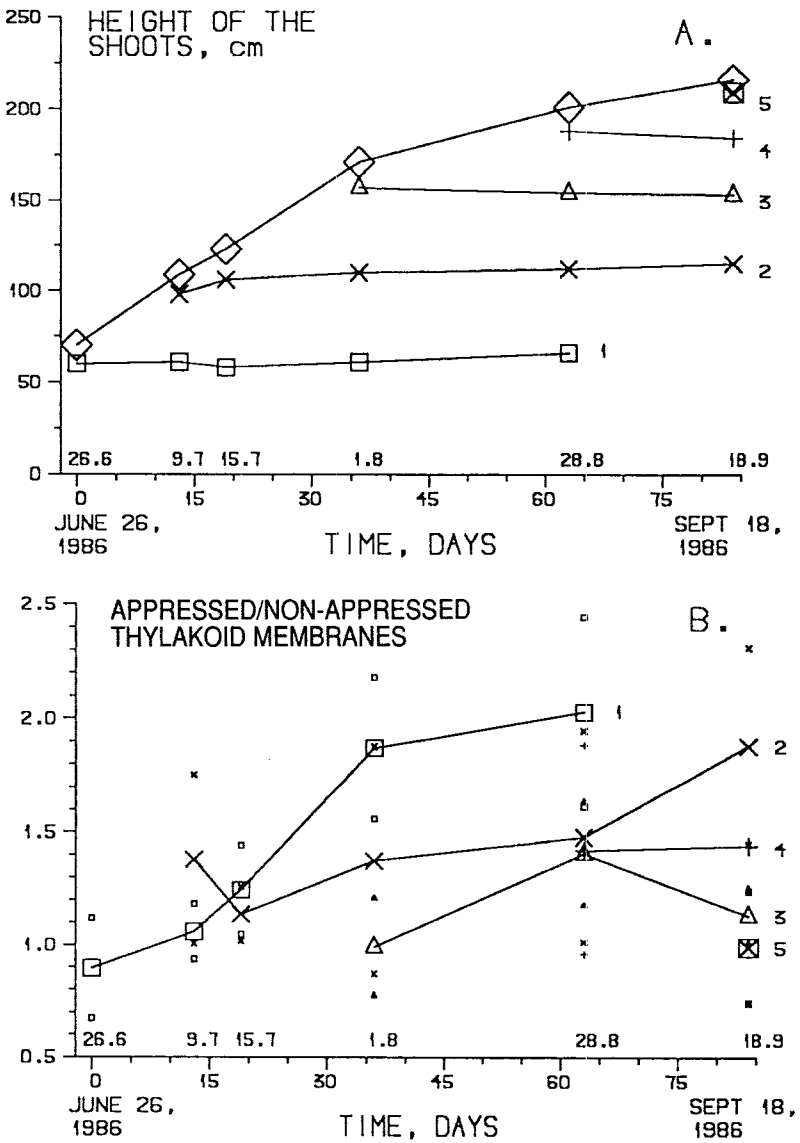


Fig. 1. A. Growth of willow shoots during the growing season. B. The ratio of total length of appressed to non-appressed thylakoid membranes vs time. The numbers 1-5 show the heights at which the samples were collected.

ultrastructure was analyzed from the electron micrographs as described by Aro *et al.*, (1986) and Vapaavuori (1986). On an average, 6 typical chloroplasts were analyzed from each sample of the 3 replicate plots.

## Results and Discussion

At all studied levels of the canopy, the ratio of the total length of appressed to

non-appressed thylakoid membranes was lowest (0.9–1.4) in the youngest leaves (Fig. 1B) that were exposed to sun (Fig. 2B). The thylakoid structure in these leaves was similar to that in plants adapted to sunny habitats or grown at high

quantum flux densities (Anderson and Osmond, 1987). At level 1 (60 cm above-ground) the ratio increased slightly until the middle of July (Fig. 1B), but remained typical of sun-exposed leaves (below 1.3). During this period, the low rates of CO<sub>2</sub>

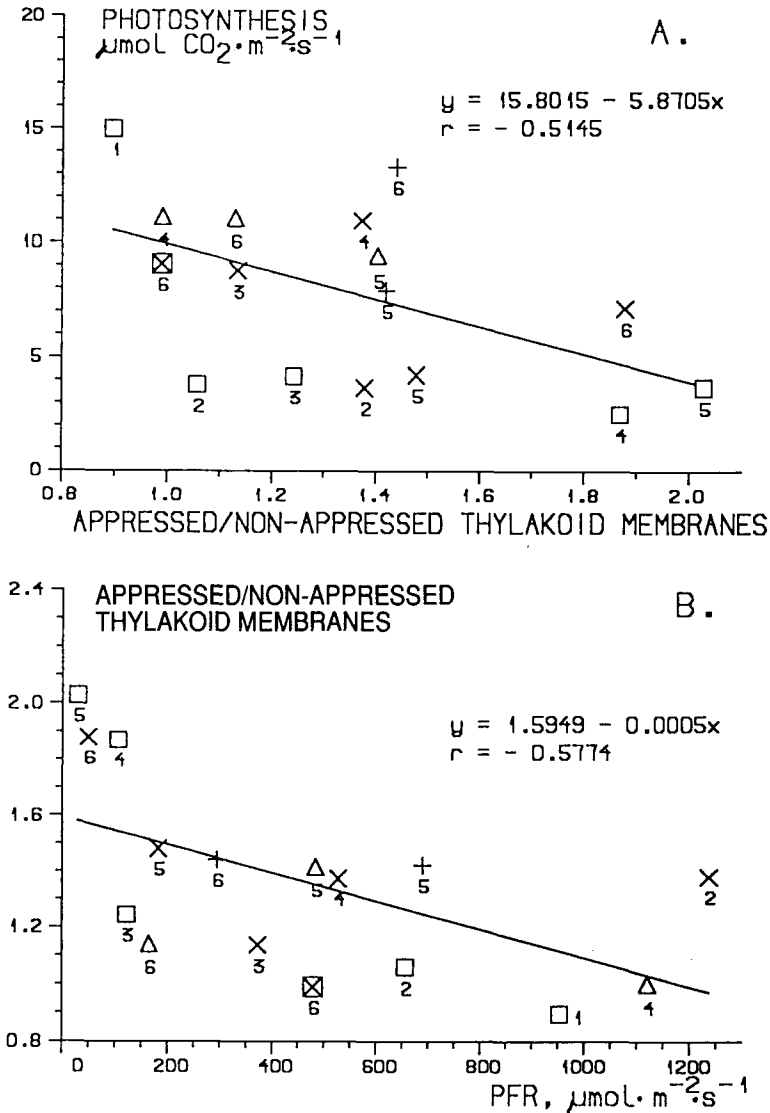


Fig. 2. A. Rate of CO<sub>2</sub> uptake vs the ratio of total length of appressed to non-appressed thylakoid membranes. B. Ratio of total length of appressed to non-appressed thylakoid membranes vs photon fluence rate (PFR). Symbols as in Fig. 1A. The numbers 1–6 refer to the sampling dates in Fig. 1.

uptake recorded (Fig. 2A) were possibly caused by decreased availability of excitation energy in the canopy and not by altered organization of thylakoid membranes. Later in the growing season, the chloroplast ultrastructure acclimated to decreased light (Fig. 2B) and the low rates of CO<sub>2</sub> uptake (Fig. 2A) were possibly caused by altered thylakoid structure typical of shade plants (Lichtenthaler *et al.* 1981). Part of this reorganization in thylakoid membranes might also be due to ageing, since the area of plastoglobuli of chloroplast area increased (data not shown), which is known to be an indication of ageing (Hudak, 1981). The pattern of thylakoid organization at level 2 (110 cm aboveground) was similar to that at level 1; only the appressed/non-appressed membrane ratio was initially somewhat higher than at level 1.

Leaves at level 3 maintained high rates of CO<sub>2</sub> uptake throughout the 7 wk period under examination (Fig. 2A), although the quantum flux density decreased markedly (Fig. 2B). The thylakoid structure was typical of sunny habitats, since the ratio of the length of appressed to non-appressed thylakoid membranes remained below 1.4 (Fig. 1B). The leaves examined from levels 4 and 5 were physiologically young and the rates of CO<sub>2</sub> uptake recorded were from intermediate to high (Fig. 2A). The ratio of the length of appressed to non-appressed thylakoid membranes was, however, quite different (Fig. 1B). One might speculate that the high ratio, 1.5, in chloroplasts at level 4 was due to the late season, as suggested by Aro *et al.* (1985). This argument is, however, not valid for the somewhat younger leaves at level 5, which had developed under similar climatic conditions but had a lower rate of CO<sub>2</sub> uptake and an appressed/non-appressed membrane ratio of about 1.

In the present study, a negative correlation was found between  $P_N$  and the ratio

of the length of appressed to non-appressed thylakoid membranes (Fig. 2A) and between the ratio of the length of appressed to non-appressed thylakoid membranes and photon fluence rate (Fig. 2B). This suggests that, in the canopy, acclimation of the thylakoid structure to decreasing photon fluence rates will lead to gradual impairment of the photosynthetic capacity.

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