

Origin identification of maritime pine stands in France using chloroplast simple-sequence repeats

Maria Margarida Ribeiro^{a,**}, Grégoire LeProvost^{b,**}, Sophie Gerber^b,
Giovanni Guiseppe Vendramin^c, Maria Anzidei^c, Stéphane Decroocq^b, Anne Marpeau^d,
Stéphanie Mariette^b and Christophe Plomion^{b,*}

^a Department of Forest Genetics and Plant Physiology, SLU, 901 83 Umeå, Sweden

^b INRA, Équipe de Génétique et Amélioration des Arbres Forestiers, BP 45, 33610 Cestas, France

^c Istituto Miglioramento Genetico Piante Forestali, CNR, Via Atto Vannucci 13, 50134 Firenze, Italy

^d Laboratoire de Chimie des Substances Végétales, Institut du Pin, Université de Bordeaux 1, 351 cours de la libération, 33405 Talence Cedex, France

(Received 20 October 2000; accepted 28 February 2001)

Abstract – Maritime pine seed-lots from north-western Iberian regions (Portugal and Galicia) were introduced in the 1950s to the south-west of France (Aquitaine region), and the stands they formed suffered considerable frost damage. In the mid 1980s, a biochemical test was developed to test the putative origin of adult stands in Aquitaine, before seeds could be distributed for commercial purposes in France. In this paper, we describe a new test employing chloroplast simple-sequence repeats (cpSSRs) to facilitate identification of stand origin based on randomisation tests. The origin of five stands of unknown origin was determined with both the cpSSR and biochemical (terpene profile analysis) tests. The results from the two tests were concordant, but the DNA-based test gave faster and more accurate results. Use of this test should help when determining the origin of maritime pine stands in the Aquitaine region of France.

cpSSR / microsatellites / terpene / origin identification / *Pinus pinaster*

Résumé – Identification de l'origine géographique des peuplements de pin maritime en France à l'aide de microsatellites chloroplastiques. Des lots de graines du nord ouest de la péninsule ibérique (Portugal et Galice) ont été introduits dans les années 1950 dans le sud-ouest de la France (Aquitaine), et les peuplements issus de ces graines ont fortement souffert des gelées. Un test variétal basé sur les marqueurs terpéniques fut développé dans les années 1980 afin d'identifier l'origine géographique des peuplements adultes en Aquitaine sur lesquels des graines étaient récoltées puis commercialisées. Dans cet article nous décrivons un nouveau test qui utilise des marqueurs microsatellites chloroplastiques (cpSSRs) et simulations pour identifier l'origine des peuplements. Une étude comparative des tests biochimique (terpènes) et cpSSR a été menée sur cinq peuplements adultes. Les résultats obtenus sont identiques, mais le test ADN s'est avéré plus rapide et plus précis. L'utilisation de ce nouveau test devrait permettre de garantir l'origine géographique des peuplements de pin maritime du sud-ouest de la France (Aquitaine).

cpSSR / microsatellites / terpène / test variétal / *Pinus pinaster*

* Correspondence and reprints

Tel.: +33 5 57 97 90 76; Fax: +33 5 57 97 90 88; e-mail: plomion@pierroton.inra.fr

** To be considered as joint first authors.

1. INTRODUCTION

Pinus pinaster is an important species in France occupying 1.4 M ha, representing 12% of the French forest area. Seeds of this species from northwestern (NW) Iberian origins (Portugal and Galicia) were introduced in the 1950s to the southwest of France (the Aquitaine region) for reforestation, since large areas of forest were burned, during and after the 2nd World War (in 1943 and 1949). Unfortunately, the stands proved to be frost-sensitive, especially during the exceptionally cold winter of 1985 in Aquitaine, when the temperature dropped to -22°C . Damage caused by frost affected about 30,000 ha of *P. pinaster* stands, and financial losses were considerable [6]. To overcome this problem and to avoid further damage, from 1986 onwards candidate stands for seed collection in the Aquitaine region had to be certified for their French origin. A diagnostic test based on a discriminant analysis of terpenes was developed by Baradat and Marpeau-Bezard [4]. This test has been routinely used to identify the origin of adult stands, by comparing their terpene profile with the profiles given by stands of known origin.

The human impact on forest species may have a negative effect in the long run, especially in the case of introduced seed material. Stands from non-indigenous origin may prove to be poorly adapted to the area, particularly if the reproductive plant material is introduced from regions with very different environmental conditions, and they may also influence adjacent native stands by pollen and seed dissemination. The establishment of forest plantations throughout the world demands increasing amounts of seed annually. Seeds are often transferred between countries, or between areas within countries, accompanied by inadequate information about their source and history. Therefore, reproductive material identification and certification has been an important issue in recent decades [16, 21].

Morphological data and biochemical markers (terpenes, isozymes and denatured proteins) have all been used for provenance identification and seed certification in forest trees (e.g. [2, 6, 11, 12]). Molecular markers based on nuclear and organelle DNA analysis have also been used recently for this purpose (e.g. [1, 7, 30, 33]).

The terpene method used for plant material certification has several constraints, such as the restricted type of plant material that can be used (cortex of completely lignified young shoots), the age of the tree for material collection (7–10 years minimum), and the limited time

that the material can be stored (eight days). In addition, for financial reasons, the terpene test uses bulk samples of tissues, instead of an individual analysis of each sample [3].

The use of DNA has several advantages over terpene analysis, because it is relatively stable, ubiquitous and convenient to analyze. DNA is present in nearly all tissues and it can be extracted using any sample taken from the plant. Plant material can be easily stored for DNA analysis under field and laboratory conditions, and only few nanograms of DNA are needed [23]. Therefore, the use of DNA markers for assessing the origin of maritime pine stands may provide an attractive alternative to other markers.

The level at which the identification is to be done is a very important factor in the choice of the marker. In our case, the aim was to identify populations of a species (*P. pinaster*) at the provenance level (NW Iberian vs. Aquitaine). The use of an appropriate genetic marker is also of great importance in order to maximize accuracy and to save time. For our study, a marker capable of clearly discriminating between provenances was required. In plants, chloroplast microsatellite regions are highly polymorphic, and they have already proved to be useful for genetic fingerprinting in different pine species (Lefort et al. [17] and references therein). Moreover, the chloroplast genome is haploid, it does not undergo recombination and it is usually paternally inherited in conifers (e.g. [8, 22, 31, 32, 36]).

In the present study we have used six chloroplast microsatellites to investigate the haplotypic composition of two populations from Spain (Galicia) and 15 populations from France. In addition, data from 12 *P. pinaster* populations from Portugal for the same cpSSR loci were included [28]. The aims of this study were to construct and optimize a cpSSR-based test in order to determine the putative origin of maritime pine forest stands in Aquitaine, and to compare the cpSSR-based test with the test based on terpene profile analysis [4].

2. MATERIALS AND METHODS

2.1. Plant material

2.1.1. Reference populations

The plant material presented in *table 1* was used as NW Iberian and French reference populations. The NW

Iberian reference material included 303 samples: 235 trees from 12 Portuguese populations (coded A-N), and 68 individuals obtained from a bulk seed-lot collected in two localities in the Galicia region of Spain (CARB and PUEN). The French reference material included 450 samples from Aquitaine: 371 trees collected from 13 natural populations (coded 1-13) along the Atlantic coast, 45 individuals obtained from a bulk seed-lot from four different populations (CEMA), and 34 “plus trees” representing the breeding population (VEC) in France.

2.1.2. Tested stands

Five adult stands of unknown origin were sampled in the Aquitaine region. Completely lignified young shoots were collected from 120 randomly chosen trees in each stand to perform the terpene analysis, but a different set of 30 was used for each repeated analysis. For the cpSSR analysis, needles from 30 trees per stand among the previously collected material were used.

2.2. CpSSR and terpene analysis

Total DNA was extracted according to the Doyle and Doyle [10] protocol with modifications described by Lerceteau and Szmidi [18] and Plomion et al. [25]. The number of individuals used per population and the type of material used for DNA extraction are shown in *table I*.

In this study, a maximum of six primers flanking pine chloroplast microsatellites were used; Pt1254, Pt15169, Pt30204, Pt36480, Pt71936 and Pt87268, designed according to sequences in the *P. thunbergii* chloroplast genome [35]. These primers were chosen since they detected a relatively high level of polymorphism in an analysis of a sub-sample of individuals.

Polymerase chain reaction (PCR) and electrophoresis were performed according to the protocol described for *P. pinaster* by Ribeiro et al. [28]. The presence of the PCR fragments was visually scored with the help of fragment size standards. To confirm the accuracy of the visual reading and to evaluate the size of the alleles, two samples of 21 and 34 individuals from the Portuguese and French populations, respectively, containing all the alleles found with the six primers were used. The size of the amplified fragments from these two samples was evaluated according to Vendramin et al. [34].

Terpene compounds were extracted from oleoresin obtained from the top-shoot cortical tissues of 30 individual trees from each of the five stands of unknown origin.

The bulk-sample analysis was performed using gas chromatography according to the method described by Baradat and Marpeau-Bezard [4]. Whenever the results obtained were inconclusive, the test was repeated up to four times, using further bulk-samples of the same size.

2.3. Differentiation statistics

In this study, for ease of presentation, the term “locus” will refer to a cpSSR site and “allele” will refer to a size variant at a given cpSSR site. Since the chloroplast genome is haploid and does not undergo recombination, the detected alleles at each locus were combined in order to derive the chloroplast haplotype of each individual.

The total among-population differentiation based on haplotype frequencies at the six cpSSR loci (using n_1 in *table I*, except CEMA and VEC) were computed for the 12 French and 12 Portuguese populations grouped together. The population differentiation among the French and Portuguese groups of populations, taken separately, was also determined (n_1 in *table I*, except CEMA and VEC). The genetic differentiation among populations was assessed by calculating θ , following Weir and Cockerham [37]. The infinite allele mutation model was preferred to the stepwise-mutation model because, in general, the θ -based estimates perform better when samples sizes are moderate or small and the number of scored loci is low [13]. The pairwise population differentiation, French against Portuguese (populations confounded) was also tested. FSTAT, a program for estimating and testing gene diversities (Goudet 2000, version 2.9.1), updated from Goudet [14] was used to obtain the genetic differentiation estimates and its variances, and to test the pairwise population differentiation.

2.4. Two-step cpSSR screening

Two steps were performed in the development of the cpSSR test for identifying stand origin. In the first, the Portuguese individuals and the n_1 individuals (*table I*) of the French populations were analyzed at the six cpSSR loci. The aim was to identify the loci that could best differentiate between the Portuguese and the French provenances.

Since nothing was known a priori about the expected distributions of the statistic described by formula (1), and because most of the haplotypes were represented by a small number of individuals (so the haplotype frequencies were subject to a rather large sampling error) [9, 19],

Table I. Plant material used in the study.

Origin	Code	n_1^b	n_2^c	Region	Alt (m)	Latitude	Long.	M ^d
Portugal	A ^a	20	20	Aveiro	30	40° 39' N	8° 36' W	A
Portugal	B ^a	20	20	Oleiros	750	39° 55' N	7° 50' W	A
Portugal	C ^a	19	19	Alcácer do Sal	20	37° 52' N	8° 30' W	A
Portugal	D ^a	20	20	Bragana	800	41° 52' N	6° 32' W	A
Portugal	E ^a	20	20	Figueira da Foz	30	40° 18' N	8° 44' W	A
Portugal	F ^a	19	19	Lousã	250	40° 09' N	8° 11' W	A
Portugal	G ^a	19	19	Monção	310	42° 04' N	8° 23' W	A
Portugal	H ^a	20	20	Mondim de Basto	480	41° 25' N	7° 55' W	A
Portugal	J ^a	20	20	Leiria	50	39° 46' N	8° 57' W	A
Portugal	L ^a	20	20	Manteigas	625	40° 24' N	7° 26' W	A
Portugal	M ^a	20	20	Montalegre	690	41° 49' N	7° 56' W	A
Portugal	N ^a	18	18	Sintra	250	38° 46' N	9° 22' W	A
Spain	CARB	0	46	Carballo	130	43° 13' N	8° 41' W	B
Spain	PUEN	0	22	Ponteareas	100	42° 10' N	8° 30' W	B
France	1	13	28	Lit-et-Mixe	30–40	44° 03' N	1° 19' W	A
France	2	11	30	St-Julien-en-Born	20	44° 06' N	1° 19' W	A
France	3	12	29	Boul. Allemands	20	44° 05' N	1° 19' W	A
France	4	12	30	Ste-Eulalie	40–50	44° 20' N	1° 14' W	A
France	5	12	27	Mimizan	35–40	44° 08' N	1° 18' W	A
France	6	13	30	Vielle St-Girons	35	43° 56' N	1° 28' W	A
France	7	8	29	Biscarosse	25–60	44° 20' N	1° 13' W	A
France	8	0	18	Biscarosse	30	44° 33' N	1° 11' W	A
France	9	13	32	Lège	15	44° 43' N	1° 12' W	A
France	10	10	25	Lacanau	10–15	45° 02' N	1° 09' W	A
France	11	12	31	Pointe de Grave	10–15	45° 34' N	1° 04' W	A
France	12	12	31	Carcans	10–15	45° 06' N	1° 09' W	A
France	13	11	31	Hourtin	25–45	45° 10' N	1° 08' W	A
France	VEC	34	34	“plus”trees	10–50	Aquitaine		A
France	CEMA	45	45	Medoc	10–15	Medoc		B

^a: Group of populations previously analyzed in [28].

^b: Number of individuals screened at six cpSSR loci.

^c: Number of individuals screened at two cpSSR loci.

^d: Type of plant material used for DNA extraction: A, needles and B, germinated embryos.

simulations had to be performed. The following formula was used to obtain the distribution of the null (H_0 : “the tested stand is of French origin”) and the alternative hypotheses (H_1 : “the tested stand belongs to the Portuguese provenance”):

$$S_j = \sum_{i=1}^n (x_{iF} - x_{ij})^2 \quad (1)$$

where n is the total number of different haplotypes found in both provenances, x_{iF} is the frequency of the i th haplotype in the French provenance and x_{ij} is the frequency of the i th haplotype in a sample from the French (to obtain H_0) or the Portuguese provenance (to obtain H_1) under the j th outcome. Resampling with replacement was performed 10,000 times ($j = 1$ to 10,000), and the

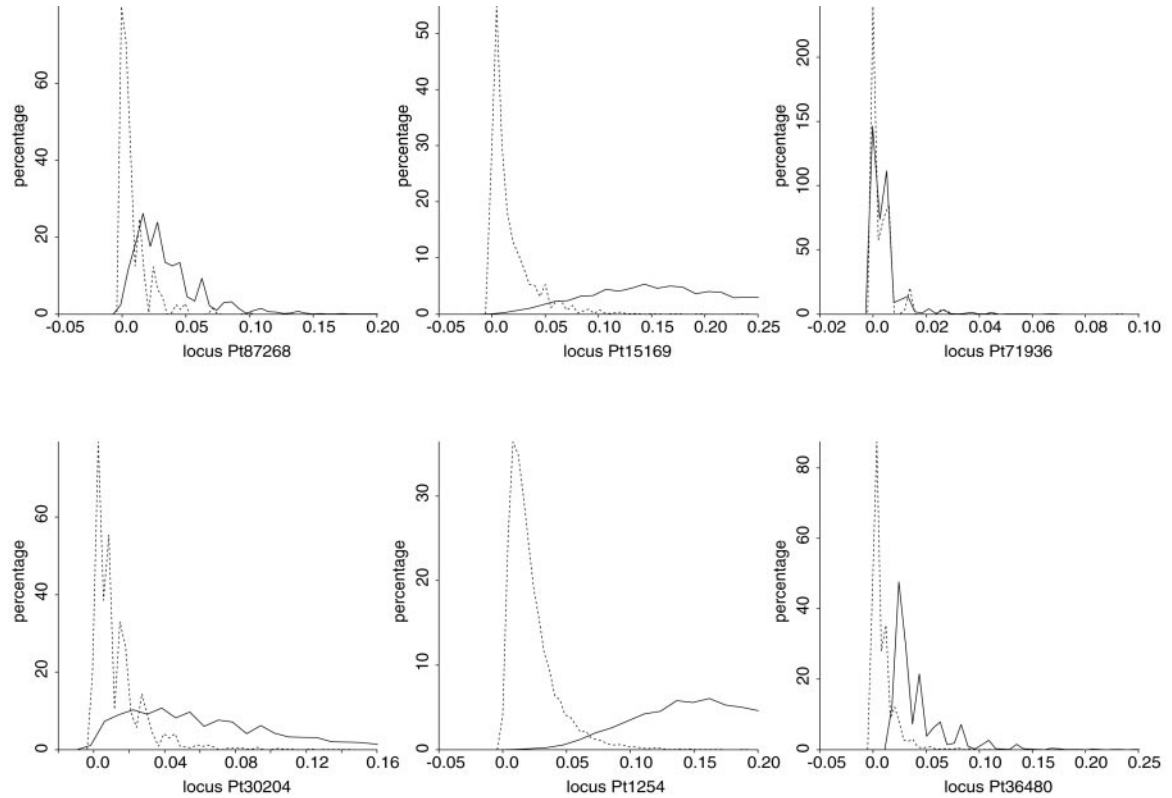


Figure 1. Density functions for the Portuguese and French reference populations (solid and dotted lines, respectively) for each locus. The data points were obtained after 10,000 simulations.

sample size was 30. The density functions, locus by locus, were obtained from the 10,000 S_j values. Random numbers were generated according to the method of Knuth proposed by Press et al. [27]

In the second step, individuals from the French, Portuguese and Spanish populations (n_2 in table I) were analyzed at the two most informative loci revealed in the first step. The complete set of data from the haplotypes derived from the two selected loci and formula (1) were used to obtain the density functions for each reference population, based on the 10,000 S_j values. The tested sample size was N ($N = 20, 30, 40$ or 50).

2.5. Testing the stands of unknown origin

In this study, for an indigenous stand the origin is the place in which the trees are growing and for a non-indigenous stand the origin is the place from which the seeds or plants were originally introduced [16].

The haplotypes of 30 individuals from each one of the five stands (designated $\lambda 1$ – $\lambda 5$) of unknown origin were recorded at the two selected loci. A statistic, S_λ , was computed for each λ stand (1 to 5), to determine the degree of similarity between a stand of unknown origin and the French provenance. The formula used was as follows:

$$S_\lambda = \sum_{i=1}^n (x_{iF} - x_{i\lambda})^2 \quad (2)$$

where n is the total number of different haplotypes found both in the French reference population and in the λ stand, x_{iF} is the frequency of the i th haplotype in the French reference population and $x_{i\lambda}$ is the frequency of the i th haplotype in the λ stand.

The critical value S_c was chosen in order to keep $\beta = 1\%$. It was considered of less importance to reject a stand of French origin when it was of French origin (type I error) than it was to accept a stand as being of French origin when it was not. The probability of accepting the null hypothesis when it was false (type II error) was kept

equal to 1%, since it was highly desirable to reject all the stands of probable NW Iberian origin. Whenever the value of the S_λ statistic was found to be smaller than the critical value, S_c , the stand was assumed to be of French origin.

3. RESULTS

3.1. Characterisation of the cpSSR loci and population diversity

A total of 25 alleles (from two to seven per locus) were detected in the 453 sampled individuals at the six cpSSR loci (samples n_1 in *table I*). The description of the alleles at each locus and the haplotypic data can be obtained upon request from the corresponding author. Exclusive alleles were found for both the Portuguese (85 base pairs (bp) for Pt1254 and 146 bp for Pt36480) and the French (164 bp for Pt87268, 112 bp for Pt15169, 142bp for Pt71936, and 142 bp for Pt30204) data sets. When all alleles were combined, 71 different haplotypes were found. While 15 (21%) of the haplotypes were common to both provenances, 39 and 17 (55% and 24%) were exclusively present in the French and Portuguese provenances, respectively. The among-population diversity obtained for the French populations was found not to be significantly different from zero: $\theta_F = 0.005 \pm 0.011$ (\pm SD). The θ value, θ_P , obtained for the Portuguese populations was very low, $\theta_P = 0.023 \pm 0.014$, but significantly different from zero. The genetic differentiation computed for both groups of populations was $\theta_T = 0.038 \pm 0.009$, and significantly different from zero. The pairwise population differentiation, French against Portuguese groups (populations confounded) was tested and found to be significant at the 0.1% probability level.

The density functions of the Spanish (Galician) and Portuguese populations overlapped (data not shown). Both functions were found to be not significantly different from each other, and the data from Portuguese and Spanish populations were merged together to obtain the density function of the NW Iberian reference population. Conversely, the density functions for the NW Iberian and the French reference populations did not overlap significantly ($\alpha = 2\%$), thus both groups could be considered divergent from each other (*table III*).

3.2. Discriminant loci and effect of the sample size

In the first screening step, the density functions for the S_j statistic were obtained for all cpSSR loci (*figure 1*), using a subset of the French and Portuguese individuals (n_1 in *table I*). The three non-discriminant loci had overlapping density functions, and three loci (Pt1254, Pt36480 and Pt15169) were found to discriminate between the two provenances (*figure 1* and *table II*).

In a second step, the haplotypes of additional individuals from both provenances (sample n_2 in *table I*) were recorded at the two most informative loci (Pt1254 and Pt36480). The power of the test ($1-\beta$) obtained for the two combined loci was higher (98.4%) than the corresponding value for each informative locus taken separately (*table II*). Since a compromise had to be found between the accuracy of the test and its costs, combinations with more than two loci were not considered. Moreover, the size of the amplified fragments from the two selected loci allowed sufficient discrimination and the amplified products from different loci could be loaded simultaneously in the same lane, which saved time and costs. *Figure 2* shows the density functions for the null and alternative hypotheses based on the two selected loci. Locus Pt15169 was excluded, because of its lower discriminatory power of the test (*table II*).

The effect of the sample size (N) used to compute the statistic S_j was tested over the type I error, when type II error was kept constant ($\beta = 1\%$). As expected, the type I error decreased with increasing sample size (*table III*). With a sample size of 30, the α value was 2% and the

Table II. Characteristics of the probability densities, using the n_1 samples described in *table I*, for each of the six cpSSR loci and for the two discriminant loci.

Locus	S_c^a	α^b (%)	$(1-\beta)^c$ (%)
Pt 87268		Non-discriminant ^d	
Pt 15169	0.0540	4.40	94.65
Pt 71936		Non-discriminant ^d	
Pt 30204		Non-discriminant ^d	
Pt 1254	0.0680	2.91	96.90
Pt36480	0.0168	0.60	95.50
Pt1254 + Pt36480	0.0785	1.40	98.40

^a: Critical value for minimized α and β values.

^b: Type I error.

^c: Power of the test.

^d: Overlapping density function curves (see *figure 1*).

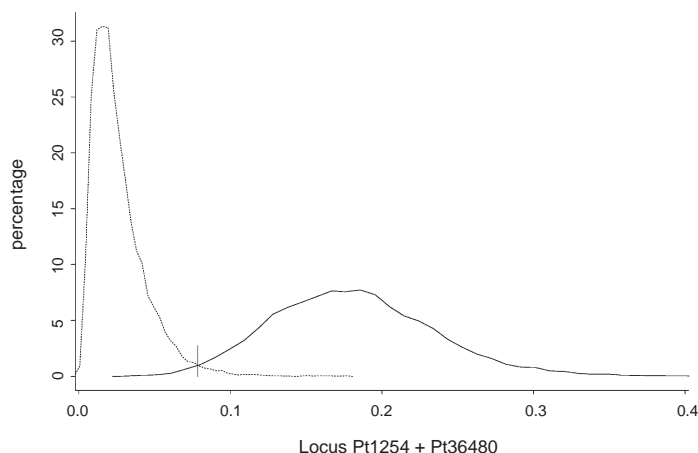


Figure 2. Density functions for the NW Iberian and French reference populations (solid and dotted lines, respectively) for the two most discriminating loci combined (Pt1254 and Pt 36480). The data points were obtained after 10,000 simulations.

Table III. Influence of the sample size (N) on the resampling procedure, with type II error of 1% and using the n_2 individuals as described in *table I*.

N	α^a (%)	S_c^b
20	15	0.065
30	2	0.075
40	0.3	0.081
50	0	0.087

^a: Type I error.

^b: Critical value with $\beta = 1\%$.

critical value was $S_c = 0.075$. Due to the time and costs needed to run larger sets of samples, and because the obtained α value was reasonable, $N = 30$ was selected as the sample size.

3.3. Testing the λ stands

The critical value $S_c = 0.075$ was compared with the statistic S_λ computed for each λ stand (*table IV*). Using the two selected loci, only one of the λ stands was found to be of NW Iberian origin, while all the remaining stands were of French origin. Furthermore, the stand of putative NW Iberian origin showed the exclusive allele found in the Portuguese populations at the locus Pt36480 (146 bp). The

terpene analysis was less conclusive. In two out of the five stands, the first biochemical analysis was insufficient to identify their putative origin. It had to be repeated up to four times until a conclusive answer was obtained and the origin of the stands was determined (*table IV*).

4. DISCUSSION

4.1. The importance of the markers used (cpSSRs)

In this study, in order to develop the test comparing the French and NW Iberian provenances it was necessary to obtain sets of data that clearly differentiated between provenances, while being insensitive to differences among populations within provenances, from the cpSSR analyses. The diversity among the Portuguese populations was found to be very low and among the French populations was found to be effectively zero. Both sets of data were considered homogeneous and used as reference populations. The tests for population differentiation showed a clear difference between the Portuguese and the French groups of populations: this means that the results obtained with the markers (cpSSRs) we used allowed the two provenances to be differentiated. The Portuguese and Galician populations together constituted the NW Iberian reference material, as they were not significantly divergent from each other. However, since the density functions for the NW Iberian and the French

Table IV. Comparison between the cpSSR (using loci Pt1254 and Pt36480) and the terpene test to discriminate between the origins of the λ stands.

Stand code	S_{λ}^a	S_c^b	cpSSR test	Terpene test			
				1st	2nd	3rd	4th
λ				1st	2nd	3rd	4th
1	0.0600	0.075	F ^c	F			
2	0.1200	0.075	NWI ^d	NWI			
3	0.0540	0.075	F	IR ^e	IR	IR	F
4	0.0600	0.075	F	F			
5	0.0057	0.075	F	IR	IR	IR	F

^a: Statistic computed for each λ stand.

^b: Critical value with $\beta = 1\%$.

^c: Stand of French origin.

^d: Stand of NW Iberian origin

^e: Inconclusive result.

reference populations did not overlap significantly, the two groups could be considered divergent from each other.

Pinus pinaster has a scattered distribution in its natural range, which probably accounts for the wide divergence found among regions. An isozyme-based study showed high levels of divergence among six populations spanning most of the distribution range of this species ($G_{ST} = 0.16$, [24]) compared with other *Pinus* species (average $G_{ST} = 0.065$, [15]). The markers used in this study (cpSSR) also revealed high levels of genetic differentiation among populations due to differences in allele size, across the range of this species [34].

In the present study, the results obtained with chloroplast microsatellites showed a homogeneous distribution of the polymorphism within groups and clear differentiation between the two groups of populations (French and NW Iberian). This could have been caused by reforestation programs that have been undertaken in most parts of the range of *P. pinaster* since the beginning of the twentieth century [5]. The use of seeds of different origins, together with gene flow, has probably obscured the divergence among populations within regions. Both of these factors could explain the homogeneity found within regions [20, 28, 29]. The current distribution of the species in Aquitaine is largely composed of artificial plantations with seeds coming from original stands located along the coast. There is strong historical and geographical evidence showing that the stands included in the French reference population (coded 1-13, *table I*) in this study are natural and of French origin (Mariette et al. [20] and references therein).

4.2. Comparison between the cpSSR and the terpene test

The results obtained with terpenes proved to be less discriminating than those obtained with the cpSSRs. In fact, with the new test, the origin of the stands was identified with no inconclusive results. In contrast, the terpene test was initially inconclusive for two out of the five tested stands and it had to be repeated up to four times before a reliable answer was obtained. Thus, use of the terpene test risks the need for repetitions, increasing both the amount of plant material and time needed to get the same information as obtained from a single cpSSR test.

In a study where the range-wide genetic structure of *P. pinaster* populations was investigated by terpene analysis, the results showed that populations from western France, Portugal and a large part of Spain form a cluster [4]. Other population genetic studies in *P. pinaster* have also indicated that the French and Portuguese populations are clustered together, using allozymes, proteins [2, 24] and, recently, mitochondrial DNA markers (Burban, unpublished results).

Conversely, the distribution of chloroplast haplotypes and the haplotypic diversity is geographically structured at the regional level in the range of different species of conifers, including *P. pinaster* as shown by both this study and by Lefort et al. [17]. In a study where ten populations of *P. pinaster* spanning the range of the species were screened using cpSSRs, an evident discontinuity between the Portuguese and French groups of populations was found [34], and the same phenomenon was observed in the present study. These findings are in

agreement with our suggestion that a marker capable of detecting differences between groups should be used to identify populations at a regional level. Therefore, the cpSSR markers used in this study were suitable for the principal purpose of our study: i.e. to determine the putative origin of *P. pinaster* stands in the Aquitaine region of France.

To our knowledge, the type of test developed here has never been used before for identifying the origin of stands. The results obtained indicate that similar tests could be used for other species where polymorphic cpSSRs or other markers are available. The approach suggested here could easily be applied to other commercial species, provided that there is a homogeneous distribution of the polymorphism within groups and clear differentiation between/among groups of populations. In addition, cpSSR primers have already been shown to cross-amplify sequences from several species, which could be very advantageous given the long time and high costs involved in identifying markers [26, 35].

4.3. Other test components

Nevertheless, the type of DNA marker used in the present study is probably not the only factor responsible for the differences found between the terpene and the cpSSR tests applied to *P. pinaster*. The terpene test uses a terpene profile analysis based on a bulk-sample of tissues from 30 different trees and a discriminant analysis. The cpSSR test we developed uses the haplotypes based on two loci from 30 different individuals and randomization tests were used for testing the putative origin of the tested stands. The higher accuracy showed by the cpSSR test used in this study, compared with the terpene test, was probably due to differences in several factors, i.e. the type of marker, the statistical analysis involved and the type of sample used (bulk vs. individual samples). The contribution of each of these factors to the relative accuracy of the tests is impossible to determine, but it is clear that for one or more of these reasons, the cpSSR test gives considerably better data.

5. CONCLUSION

In the present study the combined information from two cpSSR loci allowed a test to be designed with a type II error equal to 1%. This test is more accurate than the terpene test. The availability of a faster and more reliable

answer will be very valuable for identifying the origins of stands in the Aquitaine region of France, and also in the context of gene conservation. In addition, the type of marker used (cpSSR) could be analysed by a commercial genotyping laboratory using an automated DNA sequencer. Moreover, it is possible that similar methods could be developed for other species, for identifying the origin of seed-lots and for providing solutions to seed certification problems.

Acknowledgements: M.M. R. was supported by a PRODEP II fellowship Acção 5.2. This work was supported by a grant from the European Union (Contract IC18970200). We wish to thank Professor A.E. Szmidt and S.C. González-Martínez for the critical reading of the manuscript.

REFERENCES

- [1] Aragones A., Barrena I., Espinel S., Herran A., Ritter E., Origin of Basque populations of radiata pine inferred from RAPD data, *Ann. Sci. For.* 54 (1997) 697–703.
- [2] Bahrman N., Zivy M., Baradat P., Damerval C., Organization of the variability of abundant proteins in seven geographical origins of maritime pine (*Pinus pinaster* Ait.), *Theor. Appl. Genet.* 88 (1994) 407–411.
- [3] Baradat P.H., Marpeau A., Walter J., Terpene Markers, in: Müller-Stark G., Ziehe M. (Eds.), *Variation in European Populations of Forest Trees*, Sauerländer's Verlag, Frankfurt am Main, 1991, pp. 40–65.
- [4] Baradat P.H., Marpeau-Bezard A., *Le pin maritime, Pinus pinaster* Ait. Biologie et génétique des terpènes pour la connaissance et l'amélioration de l'espèce. Ph.D. Thesis, University of Bordeaux I, 1988.
- [5] Barbero M., Lisel R., Quezel P., Richardson D.M., Romaine F., *Pines of the Mediterranean basin*, in: Richardson D.M. (Ed.), *Ecology and biogeography of Pinus*, Cambridge University Press, Cambridge, 1998, pp. 153–170.
- [6] Boisseaux T., Influence de l'origine génétique (landaise ou ibérique) des peuplements de Pin maritime sur les dégâts causés par le froid de janvier 1985 au massif forestier aquitain. Mise au point d'un test variétal précoce utilisable pour le contrôle de lots de graines. Mémoire de l'ENITEF (Thesis), Université de Bordeaux, 1986.
- [7] Bucci G., Vendramin G.G., Delineation of genetic zones in the European Norway spruce natural range: preliminary evidence, *Mol. Ecol.* 9 (2000) 923–934.
- [8] Chiu W.-L., Sears B.B., Recombination between chloroplast DNAs does not occur in sexual crosses of *Oenothera*, *Mol. Gen. Genet.* 198 (1985) 525–528.

- [9] Crowley P.H., Resampling methods for computation-intensive data analysis in ecology and evolution, *Annu. Rev. Ecol. Syst.* 23 (1992) 405–447.
- [10] Doyle J.J., Doyle J.L., Isolation of plant DNA from fresh tissue, *Focus* 12 (1990) 13–15.
- [11] Espinel S., Aragones A., Ritter E., Performance of different provenances and of the local population of the Monterey pine (*Pinus radiata* D. Don) in northern Spain, *Ann. Sci. For.* 52 (1995) 515–519.
- [12] Falkenhagen E.R., Isozyme studies in provenance research of forest trees, *Theor. Appl. Genet.* 69 (1985) 335–347.
- [13] Gaggiotti O.E., Lange O., Rassmann K., Gliddon C., A comparison of two indirect methods for estimating average levels of gene flow using microsatellite data, *Mol. Ecol.* 8 (1999) 1513–1520.
- [14] Goudet J., FSTAT (Version 1.2): A computer program to calculate *F*-statistics, *J. Hered.* 86 (1995) 485–486.
- [15] Hamrick J., Godt M., Sherman-Broyles S., Factors influencing levels of genetic diversity in woody plant species, *New For.* 6 (1992) 95–124.
- [16] Jones N., Burley J., Seed certification, provenance nomenclature and genetic history in forestry, *Silvae Genet.* 22 (1973) 53–58.
- [17] Lefort F., Echt C., Streiff R., Vendramin G.G., Microsatellite sequences: a new generation of molecular markers for forest genetics, *For. Genet.* 6 (1999) 15–20.
- [18] Lerceteau E., Szmidi A.E., Properties of AFLP markers in inheritance and genetic diversity studies of *Pinus sylvestris* L., *Heredity* 82 (1999) 252–260.
- [19] Manly B.F.J., Randomization and Monte Carlo methods in biology, Chapman and Hall, London, 1991.
- [20] Mariette S., Chagné D., Lezier C., Pastuszka P., Raffin A., Plomion C., Kremer A., Genetic diversity within and among *Pinus pinaster* populations: comparison between AFLP and microsatellite markers, *Heredity* 86 (2001) 469–479.
- [21] Matthews J.D., Seed production and seed certification, *Unasylva* 18 (1964) 104–118.
- [22] Neale D.B., Wheeler N.C., Allard R.W., Paternal inheritance of chloroplast DNA in Douglas-fir, *Can. J. For. Res.* 16 (1986) 1152–1154.
- [23] Parker P.G., Snow A.A., Schug M.D., Booton G.C., Fuerst P.A., What molecules can tell us about populations: choosing and using a molecular marker, *Ecology* 79 (1998) 361–382.
- [24] Petit R.J., Bahrman N., Baradat P., Comparison of genetic differentiation in maritime pine (*Pinus pinaster* Ait.) estimated using isozyme, total protein and terpenic loci, *Heredity* 75 (1995) 382–389.
- [25] Plomion C., Bahrman N., Durel C.E., O'Malley D.M., Genomic mapping in *Pinus pinaster* (maritime pine) using RAPD and protein markers, *Heredity* 74 (1995) 661–668.
- [26] Powell W., Morgante M., McDevitt R., Vendramin G., Rafalski J., Polymorphic simple sequence regions in chloroplast genomes: Applications to the population genetics of pines, *Proc. Natl. Acad. Sci. USA* 99 (1995) 7759–7763.
- [27] Press W.H., Teukolsky S.A., Vetterling W.T., Flannery B.P., Numerical recipes in C: The art of scientific computing, Cambridge University Press, Cambridge, 1992.
- [28] Ribeiro M.M., Plomion C., Petit R., Vendramin G.G., Szmidi A.E., Variation of chloroplast simple-sequence repeats in Portuguese maritime pine (*Pinus pinaster* Ait.), *Theor. Appl. Genet.* 102 (2001) 97–103.
- [29] Salvador L., Alía R., Agúndez D., Gil L., Genetic variation and migration pathways of maritime pine (*Pinus pinaster* Ait) in the Iberian peninsula, *Theor. Appl. Genet.* 100 (2000) 89–95.
- [30] Sinclair W.T., Morman J.D., Ennos R.A., Multiple origins for Scots pine (*Pinus sylvestris* L) in Scotland: evidence from mitochondrial DNA variation, *Heredity* 80 (1998) 233–240.
- [31] Stine M., Keathley D.E., Paternal inheritance of plastids in Engelmann spruce × blue spruce hybrids, *J. Hered.* 81 (1990) 443–446.
- [32] Szmidi A.E., Alden T., Hällgren J.-E., Paternal inheritance of chloroplast DNA in *Larix*, *Plant Mol. Biol.* 9 (1987) 59–64.
- [33] Szmidi A.E., El-Kassaby Y.A., Sigurgeirsson A., Alden T., Lindgren D., Hällgren J.-E., Classifying seedlots of *Picea sitchensis* and *P. glauca* in zones of introgression using restriction analysis of chloroplast DNA, *Theor. Appl. Genet.* 76 (1988) 841–845.
- [34] Vendramin G.G., Anzidei M., Madaghiale A., Bucci G., Distribution of genetic diversity in *Pinus pinaster* Ait. as revealed by chloroplast microsatellites, *Theor. Appl. Genet.* 97 (1998) 456–463.
- [35] Vendramin G.G., Lelli L., Rossi P., Morgante M., A set of primers for the amplification of 20 chloroplast microsatellites in *Pinaceae*, *Mol. Ecol.* 5 (1996) 595–598.
- [36] Wagner D.B., Govindaraju D.R., Yeatman C.W., Pitel J.A., Paternal chloroplast DNA inheritance in a diallel cross of Jack pine (*Pinus banksiana* Lamb.), *J. Hered.* 80 (1989) 483–485.
- [37] Weir B.S., Cockerham C.C., Estimating *F*-statistics for the analysis of population structure, *Evolution* 38 (1984) 1358–1370.