

# Mesophyll conductance to CO<sub>2</sub> and leaf morphological characteristics under drought stress during *Quercus ilex* L. resprouting

Isabel FLECK<sup>1\*,\*\*</sup>, Karen PEÑA-ROJAS<sup>2,\*\*</sup>, Xavier ARANDA<sup>1,3</sup>

<sup>1</sup> Departament de Biologia Vegetal, Facultat Biologia, Universitat de Barcelona, Diagonal 645, 08028 Barcelona, Spain

<sup>2</sup> Present address: Facultad de Ingeniería Forestal, Universidad de Chile Santa Rosa, 11315, La Pintana, Santiago, Chile

<sup>3</sup> Present address: Departament de Tecnologia Hortícola, Institut de Recerca i Tecnologia Agroalimentàries (IRTA), Torre Marimón, Caldes de Montbui, 08140 Barcelona, Spain

(Received 15 June 2009; accepted 13 August 2009)

## Keywords:

carbon isotopic composition /  
leaf structure /  
mesophyll conductance /  
photosynthesis limitants /  
*Quercus ilex*

## Mots-clés :

composition isotopique du carbone /  
structure des feuilles /  
conductance mésophyllienne /  
limitation de la photosynthèse /  
*Quercus ilex*

## Abstract

• *Quercus ilex* L., the dominant species in Mediterranean forests and one with a great capacity for resprouting after disturbances, is threatened by the expected increase in fire frequency and drought associated with climate change.

• The aim of this study was to determine the contribution of photosynthesis limitants, especially mesophyll conductance ( $g_{mes}$ ) during this species' resprouting and under summer drought.

• Resprouts showed 5.3-fold increased  $g_{mes}$  and 3.8-fold increased stomatal conductance ( $g_s$ ) at mid-day with respect to leaves of undisturbed individuals. With increased drought, structural changes (decreased density and increased thickness) in resprouts contributed to the observed higher photosynthesis and increased  $g_{mes}$ . However,  $g_{mes}$  only partially depended on leaf structure, and was also under physiological control. Resprouts also showed lower non-stomatal limitations (around 50% higher carboxylation velocity ( $V_{c,max}$ ) and capacity for ribulose-1,5-bisphosphate regeneration ( $J_{max}$ )). A significant contribution of  $g_{mes}$  to leaf carbon isotope discrimination values was observed.

•  $g_{mes}$  exhibits a dominant role in photosynthesis limitation in *Q. ilex* and is regulated by factors other than morphology. During resprouting after disturbances, greater capacity to withstand drought, as evidenced by higher  $g_{mes}$ ,  $g_s$  and lower non-stomatal limitants, enables increased photosynthesis and rapid growth.

## Résumé – Conductance mésophyllienne pour le CO<sub>2</sub> et caractéristiques morphologiques des feuilles sous stress hydrique pendant la repousse de *Quercus ilex* L.

• *Quercus ilex* L., l'espèce dominante dans les forêts méditerranéennes qui a une grande capacité de rejets après des perturbations, est menacée par l'augmentation prévue de la fréquence des incendies et de la sécheresse associées au changement climatique.

• Le but de cette étude était de déterminer, chez cette espèce, la contribution des limitations de la photosynthèse, en particulier de la conductance du mésophylle ( $g_{mes}$ ) au cours de la repousse et sous sécheresse estivale.

• Les feuilles des rejets ont présenté une conductance mésophyllienne ( $g_{mes}$ ) 5,3 fois plus élevée et une conductance stomatique ( $g_s$ ) à midi 3,8 fois plus élevée par rapport aux feuilles d'arbres non perturbés. Avec l'accroissement de la sécheresse, les changements de structures (diminution de la densité et épaisseur accrue) dans les rejets ont contribué à augmenter la photosynthèse et à accroître  $g_{mes}$ . Toutefois,  $g_{mes}$  dépendait partiellement de la structure des feuilles, et était également sous contrôle physiologique. Les rejets ont aussi montré une absence de limitation stomatique (vitesse de carboxylation ( $V_{c,max}$ ) environ 50% plus élevée et une capacité de régénération pour le ribulose-1,5-bisphosphate ( $J_{max}$ )). Une contribution significative de  $g_{mes}$  à la discrimination isotopique du carbone dans les feuilles a été observée.

• La conductance mésophyllienne ( $g_{mes}$ ) a présenté un rôle dominant dans la limitation de la photosynthèse chez *Q. ilex* et est régulée par des facteurs autres que la morphologie. Au cours de la repousse après des perturbations, une plus grande capacité à résister à la sécheresse, mise en évidence par une  $g_{mes}$  et une  $g_s$  plus élevées, et une diminution des limitations non stomatiques, permettent une augmentation de la photosynthèse et une croissance rapide.

\* Corresponding author: ifleck@ub.edu

\*\* These authors contributed equally to this work.

## 1. INTRODUCTION

Holm-oak (*Quercus ilex* L.) is a deep-rooted, evergreen dominant species in Mediterranean forests which has a great capacity for resprouting after fire, clear-cut, grazing or other disturbances. Resprouts after any of these events show decreased shoot/root ratios, which makes more water and nutrients available to the shoot than in the original plants and favours photosynthesis stimulation and rapid growth (Fleck et al., 1998). *Q. ilex* is exposed to multiple environmental stress factors such as drought, heat shock, chilling, nutrient deprivation and high light stress amongst others. Increased probability of drought, heat and rising atmospheric CO<sub>2</sub> concentration during the coming decades may be particularly important in the Mediterranean basin (Christensen et al., 2007). Moreover, the expected increased risk of uncontrolled fire episodes could lead to the exhaustion of several species, generating a decline in their resprouting capacity and recovery.

There has been a long-standing controversy as to whether drought limits photosynthesis by stomatal closure, metabolic impairment or through diffusive resistances (Lawlor and Tezara, 2009). Of these resistances, CO<sub>2</sub> transfer conductance inside the leaf or mesophyll conductance ( $g_{mes}$ ) is considered relevant to photosynthesis (Flexas et al., 2008). Metabolic photosynthesis limitations (e.g. injuries to photosynthetic biochemistry and photochemistry) during drought may only be apparent: drought produces low  $g_s$ , closely related to  $g_{mes}$ , resulting in a decreased availability of CO<sub>2</sub> in the chloroplast, which down-regulates the biochemical machinery of photosynthesis.  $g_{mes}$  can be affected by leaf morphology (Terashima et al., 2001); in fact, previous results of our group (Peña-Rojas et al., 2005) related changes in  $g_{mes}$  in nursery-grown holm-oak plants submitted to water stress to variations in leaf anatomy and gas-exchange parameters.

Carbon isotope discrimination ( $\Delta^{13}C$ ) is largely due to Rubisco (which discriminates against <sup>13</sup>C during RuBP carboxylation), with the amount of discrimination depending on the ratio of CO<sub>2</sub> partial pressure at the carboxylation site ( $C_c$ ) to CO<sub>2</sub> partial pressure in the surrounding air ( $C_a$ ), which is affected by  $g_s$  and  $g_{mes}$  (Farquhar et al., 1989). As described above, morphological characteristics can affect internal resistances; thus, leaf thickness and leaf density as components of the leaf mass per area parameter (LMA) (Niinemets, 1999), can be an important source of variation in  $\Delta^{13}C$ .

The aim of this study was to characterize the photosynthetic limitants during holm oak regrowth after a clear-cut, and especially the contribution of mesophyll conductance ( $g_{mes}$ ) under drought conditions. Two kinds of resprout were used for this study, which differed in their cutting season: winter, when plants had a high availability of stored underground reserves, and summer, when part of the stored reserves had already been remobilized and used to support early growth. Other aims were to relate the morphological characteristics of resprouts to the observed  $g_{mes}$  and to examine the effect of  $g_{mes}$  on carbon isotope discrimination ( $\Delta^{13}C$ ) values. The characterization of the photosynthetic and growth limitations during *Q. ilex* resprouting after disturbances would help us to establish the adaptation

capacity of this plant in the context of global change and biodiversity conservation in Mediterranean forests.

## 2. MATERIALS AND METHODS

### 2.1. Experimental site and plant material

The study was carried out at Can Coll, Serra de Collserola forest, Barcelona, Spain; 41° 28' 28" N, 2° 7' 32" E. A plot (400 × 280 m) at altitude of 140 m and oriented N-NE was selected. The climate is Mediterranean, with cold winters, cool and wet springs and autumns, and hot dry summers (Tab. I). The 35-year old forest is dominated by *Quercus ilex* and *Pinus halepensis*. In February, 25 *Quercus ilex* plants were selected (5.9 ± 0.3 cm mean diameter at breast height (DBH), 4.7 ± 0.2 m mean height, 1.4 ± 0.2 kg mean leaf biomass) and the shoots of 10 randomly selected plants were completely excised 15 cm above soil level. Resprouts (R) after this date were designated as RW (winter resprouts). In August, 10 more plants were completely excised and resprouts after this were designated as RS (summer resprouts). Five plants were kept undisturbed, as controls (C) of the clear-cut site. Leaf gas exchange and chlorophyll fluorescence were measured in fully expanded leaves of the same age: in the first winter (W; February–March), only controls and RW leaves were analysed, as RS had resprouted badly in the autumn; in the subsequent summer (S; July–August), all treatments could be analysed. Samples were collected for <sup>13</sup>C composition ( $\delta^{13}C$ ), leaf mass per area (LMA), leaf density (D) and leaf thickness (T) determinations.

### 2.2. Leaf gas exchange

A portable gas exchange system LI-6200 (Li-Cor Inc., Lincoln, NE, USA) was used for punctual measurements at midday on nine attached, fully expanded, current-year leaves per treatment, season and leaf orientation. Leaf cuvette conditions differed according to the season (Tab. I). Results were expressed per leaf-projected area (LA), obtained with an Epson GT5000 scanner and processed using image analyser software. In each season, ten CO<sub>2</sub> response curves of CO<sub>2</sub> assimilation vs. intercellular CO<sub>2</sub> concentration ( $A/C_i$ ) were obtained per treatment on attached leaves with a LI-6400 instrument (Li-COR, Lincoln, Nebraska, USA). Leaf cuvette conditions were established according to the season and time of the day to reproduce a typical day in every season.

For  $A/C_i$  curves, PPFD was established as 600  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , which is saturating under these conditions (Peña-Rojas et al., 2004); a range of ambient CO<sub>2</sub> concentration ( $C_a$ ) from 50 to 800  $\mu\text{mol mol}^{-1}$  was covered. Analyses of the curves permitted the determination of:  $A_{max}$ , net photosynthesis at saturating  $C_i$  and PPFD;  $V_{c,max}$ , maximum carboxylation velocity of Rubisco;  $J_{max}$ , maximum electron transport contributing to RuBP regeneration;  $l_s$ , stomatal limitation to  $A$  ( $l_s(\%) = 100 \times (1 - (A/A_{sat}))$ );  $A_{sat}$ , net photosynthesis at saturating light and  $C_i = 350 \mu\text{mol mol}^{-1}$ .

To assess the effect of heterogeneous stomatal conductance across the leaf surface, steady-state chlorophyll fluorescence was measured in six spots of 27 leaves of the same plants used in the experiment. Water potential ( $\Psi$ ) of the same leaves was also obtained with a Scholander-type pressure pump (Soil Moisture 3005, Soilmoisture Equipment Corp., Goleta, CA, USA). The coefficient of variation of  $\Phi_{PSII}$  (see below) was not statistically higher than system repetitiveness (around 9%), indicating the absence of patchiness, and did not correlate with  $\Psi$ .

**Table I.** Climatological data recorded at the forest site during the gas-exchange measurements of the different treatments (control, C; winter sprouts, RW; summer sprouts, RS); data are the mean  $\pm$ SE of all measurements.

	Leaf Orientation	February–March		July–August		
		C	RW	C	RW	RS
PPFD ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ )	North	416 $\pm$ 150	508 $\pm$ 117	1393 $\pm$ 126	1559 $\pm$ 109	1708 $\pm$ 57
	South	1134 $\pm$ 98	1047 $\pm$ 171	1463 $\pm$ 136	1239 $\pm$ 159	1669 $\pm$ 34
VPD (KPa)	North	2.2 $\pm$ 0.1	1.9 $\pm$ 0.1	4.5 $\pm$ 0.2	4.2 $\pm$ 0.2	4.0 $\pm$ 0.3
	South	2.2 $\pm$ 0.2	2.0 $\pm$ 0.1	4.5 $\pm$ 0.3	3.7 $\pm$ 0.3	3.8 $\pm$ 0.2
Air temperature ( $^{\circ}\text{C}$ )	North	22.2 $\pm$ 0.36	21.9 $\pm$ 0.4	36.2 $\pm$ 0.4	35.6 $\pm$ 0.4	35.6 $\pm$ 0.8
	South	22.4 $\pm$ 0.3	22.8 $\pm$ 0.7	35.9 $\pm$ 0.7	34.4 $\pm$ 0.8	35.2 $\pm$ 0.6
Leaf temperature ( $^{\circ}\text{C}$ )	North	22.9 $\pm$ 0.8	22.4 $\pm$ 0.6	39.3 $\pm$ 1.1	38.6 $\pm$ 0.6	38.9 $\pm$ 0.8
	South	24.3 $\pm$ 0.4	24.8 $\pm$ 1.1	39.7 $\pm$ 0.9	35.6 $\pm$ 0.7	37.6 $\pm$ 0.6

### 2.3. Chlorophyll fluorescence and calculation of mesophyll conductance and CO<sub>2</sub> concentration in the chloroplast

Chlorophyll fluorescence parameters were quantified with a portable modulated fluorometer (Mini-PAM Photosynthesis Yield Analyzer, Walz, Effeltrich, Germany) on the same leaves used for gas-exchange measurements. Fluorescence parameters ( $F_m$ ,  $F'_m$ ,  $F_o$  and  $F_v$ ), photochemical PSII efficiency ( $\Phi_{\text{PSII}}$ ) and the maximum quantum yield at midday ( $F_v/F_m$ ) were determined as described (Fleck et al., 1998). Non-photochemical quenching (NPQ) was calculated using the Stern-Volmer equation:  $\text{NPQ} = ((F_m/F'_m) - 1)$ . Adaptation took at least 20 min, after which  $F_v/F_m$  values reached about 95% of the pre-dawn values in *Q. ilex* (Fleck et al., 1998).

Mesophyll conductance ( $g_{\text{mes}}$ ) and CO<sub>2</sub> concentration in the chloroplast ( $C_c$ ) were calculated from combined gas-exchange (LiCor 6400) and chlorophyll fluorescence (Mini-PAM) measurements, as described by Epron et al. (1995), and Galmés et al. (2007), except for respiration, which was calculated in the same leaves at the end of an A/PPFD curve after a five min acclimatisation to darkness. Galmés et al. (2007) showed that this method yields equivalent results to the “constant  $J$ ” method (Harley et al., 1992), which makes no *a priori* assumption about the relationship between electron transport and fluorescence. Moreover, Flexas et al. (2007) demonstrated that both methods gave results that were comparable to Ethier and Livingston’s findings (2004), which did not rely on fluorescence measurements and to calculations by carbon isotope discrimination.

The rate of electron transport (ETR) was calculated as  $\text{ETR} = \Phi_{\text{PSII}} \times \text{PPDF} \times 0.5 \times 0.82$ , where 0.5 is a factor that assumes equal distribution of energy between the two photosystems and 0.82 is the light absorbance we obtained on *Q. ilex* leaves using an integrating sphere. According to the model of Epron et al. (1995), ETR can be divided into two component fractions,  $\text{ETR}_A + \text{ETR}_p$ , used for CO<sub>2</sub> assimilation and for photorespiration, respectively. To calculate  $C_c$ , we used  $S = (\text{ETR}_A/\text{ETR}_p)/(C_c/O)$  (Laing et al., 1974), where  $S$  is the specificity factor of Rubisco and  $O$  is the oxygen model fraction in the air. We used a value of  $S = 93.3 \text{ mol mol}^{-1}$  (Balaguer et al., 1996) that was corrected for leaf temperature according to Brooks and Farquhar (1985). The ratio between mesophyll conductance to CO<sub>2</sub> and stomatal conductance ( $g_{\text{mes}}/g_s$ ) was calculated at midday.

### 2.4. Leaf carbon isotope composition

Sixteen leaves per six plants per treatment and season were collected, oven-dried at 65  $^{\circ}\text{C}$  to constant dry weight and ground in a

Mixer-Mill 8000 (Spex) in vials with tungsten carbide balls. Water-soluble extracts were prepared as follows: 2 g of dry material per plant were suspended in 25 mL water (3 replicates per plant) and were heated to 100  $^{\circ}\text{C}$  for 15 min; after cooling to room temperature, samples were filtered (Whatman nr. 1), stored at  $-40^{\circ}\text{C}$  and lyophilized. Approximately 4 mg of the lyophilized water-soluble extract (WSE) and 4 mg of dry mass ( $M_d$ ) were fed into a gas chromatograph (Carlo-Erba NA1500 Series II elemental analyser, CE Elantech, Inc., Lakewood, NJ, USA), connected on-line to an isotope ratio mass spectrometer (IRMS, Finnigan, Delta S; Thermo Finnigan, San Jose, CA, USA) for  $\delta^{13}\text{C}$  determination.  $\delta^{13}\text{C}$  values were determined using a standard calibrated against Pee Dee Belemnite (PDB) carbonate and used to estimate carbon isotope discrimination ( $\Delta^{13}\text{C}$ ) as:  $\Delta^{13}\text{C} = 1000 \cdot (\delta_a - \delta_p)/(1 + \delta_p)$ , where  $\delta_a$  and  $\delta_p$  are values for air ( $-7.8\text{‰}$ ) and the plant, respectively (Farquhar et al., 1989).

### 2.5. Relative water content and leaf biomass parameters

Relative water content (RWC) was measured at midday in five young leaves of five plants per treatment. RWC was calculated as  $[(M_f - M_d)/(M_{fs} - M_d) \times 100]$ , with  $M_f$  being plant fresh mass;  $M_{fs}$ , plant fresh saturated mass (after rehydrating samples for 24 h in the dark); and  $M_d$ , plant dry mass (after oven-drying samples at 65  $^{\circ}\text{C}$  until constant weight). Leaf mass per area, LMA, was determined ( $M_d/LA$ ), and its components ( $M_f/LA$ ) and  $[(M_d/M_f) \times 100]$ , as indicators of leaf thickness ( $T$ ) and leaf density ( $D$ ), respectively (Niinemets, 1999), were calculated on the same plants as for gas-exchange measurements (30 leaves per treatment) in winter and summer.

### 2.6. Statistical analyses

All statistical procedures were carried out through the SPSS in Windows (v. 11.0, SPSS Inc., Chicago, IL, USA). Analysis of variance (ANOVA) tested the main effects and interactions, against appropriate error terms. Main factors per treatment and season for all variables were analysed. Leaf orientation was included for gas exchange and chlorophyll fluorescence analyses. The kinds of material analysed (WSE, DM) were included in the analyses of parameters derived from leaf  $\delta^{13}\text{C}$ . The *post-hoc* Duncan test was applied where suitable. Differences were considered significant at  $p \leq 0.05$ . Only statistically significant differences are described in the Results and Discussion that follow.

**Table II.** Midday values of net photosynthesis ( $A$ ), stomatal conductance ( $g_s$ ), intercellular  $\text{CO}_2$  concentration ( $C_i$ ), instantaneous water use efficiency ( $\text{WUE}_i = A/g_s$ ), relative water content (RWC), PSII efficiency ( $\Phi_{\text{PSII}}$ ), non-photochemical quenching of fluorescence (NPQ) and potential quantum yield of PSII at midday ( $F_v/F_m$ ). Data are presented according to treatment (control, C; winter resprouts, RW; summer resprouts, RS), leaf orientation (north, south) and season (winter, summer). Values are mean  $\pm$ SE of nine replicates. Significant differences across rows or columns ( $p \leq 0.05$ ) are indicated by different letters (treatment (a, b, c), season (A, B) and leaf orientation ( $\alpha$ ,  $\beta$ )).

	Season	Leaf orientation	Treatments		
			C	RW	RS
$A$ ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ )	Winter	North	$4.47 \pm 0.32^{aA\alpha}$	$4.84 \pm 0.38^{aA\alpha}$	—
		South	$5.67 \pm 0.66^{aA\beta}$	$6.82 \pm 0.77^{aA\beta}$	—
	Summer	North	$1.20 \pm 0.20^{aB\alpha}$	$3.28 \pm 0.39^{bB\alpha}$	$4.03 \pm 0.81^{b\alpha}$
		South	$0.87 \pm 0.09^{aB\beta}$	$4.55 \pm 0.69^{bB\beta}$	$5.35 \pm 0.33^{b\beta}$
$g_s$ ( $\text{mmol m}^{-2} \text{s}^{-1}$ )	Winter	North	$272.1 \pm 34.3^{aA\alpha}$	$300.6 \pm 27.6^{aA\alpha}$	—
		South	$253.1 \pm 46.6^{aA\alpha}$	$263.7 \pm 40.0^{aA\alpha}$	—
	Summer	North	$41.8 \pm 2.9^{aB\alpha}$	$123.9 \pm 12.1^{bB\alpha}$	$144.0 \pm 12.2^{b\alpha}$
		South	$37.6 \pm 5.8^{aB\alpha}$	$144.7 \pm 21.9^{bB\alpha}$	$165.3 \pm 22.1^{b\alpha}$
$C_i$ ( $\mu\text{mol mol}^{-1}$ )	Winter	North	$291 \pm 10^{aA\alpha}$	$278 \pm 6^{aA\alpha}$	—
		South	$241 \pm 13^{aA\beta}$	$240 \pm 11^{aA\beta}$	—
	Summer	North	$276 \pm 8^{aA\alpha}$	$248 \pm 13^{aB\alpha}$	$247 \pm 8^{a\alpha}$
		South	$283 \pm 4^{aB\alpha}$	$254 \pm 12^{bA\alpha}$	$240 \pm 4^{b\alpha}$
$\text{WUE}_i$ ( $\mu\text{mol mmol}^{-1}$ )	Winter	North	$19.78 \pm 3.4^{aA\alpha}$	$17.95 \pm 2.0^{aA\alpha}$	—
		South	$26.13 \pm 4.7^{aA\beta}$	$29.44 \pm 4.0^{aA\beta}$	—
	Summer	North	$28.58 \pm 3.8^{aB\alpha}$	$26.82 \pm 2.1^{aB\alpha}$	$28.35 \pm 5.1^{a\alpha}$
		South	$24.14 \pm 2.6^{aB\beta}$	$32.63 \pm 5.6^{bA\beta}$	$33.95 \pm 3.1^{b\beta}$
RWC (%)	Winter	North	$75.5 \pm 2.8^{aA\alpha}$	$85.3 \pm 1.4^{bA\alpha}$	—
		South	$73.7 \pm 1.5^{aA\alpha}$	$83.0 \pm 1.7^{bA\alpha}$	—
	Summer	North	$67.8 \pm 2.2^{aB\alpha}$	$78.6 \pm 2.1^{bA\alpha}$	$77.6 \pm 1.9^{b\alpha}$
		South	$65.2 \pm 1.2^{aB\alpha}$	$77.9 \pm 1.1^{bA\alpha}$	$77.7 \pm 1.4^{b\alpha}$
$\Phi_{\text{PSII}}$	Winter	North	$0.71 \pm 0.01^{aB\beta}$	$0.64 \pm 0.03^{aB\alpha}$	—
		South	$0.52 \pm 0.07^{aB\alpha}$	$0.56 \pm 0.04^{aB\alpha}$	—
	Summer	North	$0.09 \pm 0.02^{aA\alpha}$	$0.09 \pm 0.01^{aA\alpha}$	$0.08 \pm 0.03^{a\alpha}$
		South	$0.04 \pm 0.02^{aA\alpha}$	$0.086 \pm 0.01^{bA\alpha}$	$0.08 \pm 0.01^{b\alpha}$
NPQ	Winter	North	$0.17 \pm 0.05^{aA\alpha}$	$0.26 \pm 0.09^{aA\alpha}$	—
		South	$0.83 \pm 0.25^{aA\beta}$	$0.60 \pm 0.13^{aA\beta}$	—
	Summer	North	$2.36 \pm 0.29^{aB\alpha}$	$1.88 \pm 0.20^{aB\alpha}$	$1.93 \pm 0.04^{a\alpha}$
		South	$2.72 \pm 0.22^{bB\alpha}$	$2.01 \pm 0.18^{aB\alpha}$	$2.32 \pm 0.26^{a\alpha}$
$F_v/F_m$	Winter	North	$0.77 \pm 0.01^{aB\alpha}$	$0.75 \pm 0.02^{aB\alpha}$	—
		South	$0.76 \pm 0.01^{aB\alpha}$	$0.75 \pm 0.01^{aB\alpha}$	—
	Summer	North	$0.68 \pm 0.03^{aA\alpha}$	$0.68 \pm 0.01^{aA\alpha}$	$0.70 \pm 0.02^{a\alpha}$
		South	$0.65 \pm 0.01^{aA\alpha}$	$0.67 \pm 0.03^{aA\alpha}$	$0.66 \pm 0.03^{a\alpha}$

### 3. RESULTS

Although the two kinds of resprout used for this study differed in their cutting season, the only difference found between them was the time the resprouts took to appear: RW resprouted in the following spring, 2–3 months after cutting, whereas RS resprouted badly in the autumn and were suitable for photosynthesis measurements only from the next spring onwards (7–8 months after cutting). Since from this moment on they showed no difference from RW plants in the parameters anal-

ysed, all kinds of resprouts will be considered as R in the Discussion section.

#### 3.1. Leaf gas exchange and chlorophyll fluorescence

$A$ ,  $g_s$  and instantaneous water use efficiency ( $\text{WUE}_i = A/g_s$ ) at midday (Tab. II) showed no difference between treatments in winter, whereas in summer, resprouts gave higher values than C. Declines in  $A$  and  $g_s$  between winter and summer were observed for all treatments, but were more pronounced in C,



**Table III.** Net CO<sub>2</sub> assimilation at saturating C<sub>i</sub> and light (A<sub>max</sub>), maximum carboxylation velocity of Rubisco (V<sub>c,max</sub>), maximum potential rate of electron transport contributing to RuBP regeneration (J<sub>max</sub>) and stomatal limitation (l<sub>s</sub>) from A/C<sub>i</sub> curves for the different treatments (control, C; winter sprouts, RW; summer sprouts, RS) and season (winter, summer). In summer, the time of day was also considered. Each value represents the mean ±SE of ten replicates. Significant differences across rows or columns (*p* ≤ 0.05) are indicated by different letters (treatment (a, b, c), season (A, B) and time of the day (α, β, γ)).

	Season	Time of the day	Treatments			
			C	RW	RS	
A <sub>max</sub> (μmol m <sup>-2</sup> s <sup>-1</sup> )	Winter	Midday	6.6 ± 0.5 <sup>aB</sup>	7.2 ± 0.3 <sup>aB</sup>	—	
		Morning	2.7 ± 0.3 <sup>aβ</sup>	6.1 ± 0.6 <sup>bβ</sup>	6.5 ± 0.9 <sup>bβ</sup>	
	Summer	Midday	1.1 ± 0.2 <sup>αA</sup>	3.4 ± 0.5 <sup>bαA</sup>	4.8 ± 1.3 <sup>bα</sup>	
		Evening	1.1 ± 0.3 <sup>αα</sup>	3.6 ± 0.6 <sup>bα</sup>	3.4 ± 0.2 <sup>bα</sup>	
	V <sub>c,max</sub> (μmol m <sup>-2</sup> s <sup>-1</sup> )	Winter	Midday	30.7 ± 2.8 <sup>aB</sup>	31.8 ± 1.6 <sup>αA</sup>	—
			Morning	11.8 ± 2.1 <sup>αα</sup>	26.6 ± 3.5 <sup>bα</sup>	27.3 ± 4.3 <sup>bα</sup>
Summer		Midday	13.5 ± 4.0 <sup>ααA</sup>	29.9 ± 2.3 <sup>bαA</sup>	25.1 ± 5.7 <sup>bα</sup>	
		Evening	11.8 ± 2.4 <sup>αα</sup>	27.8 ± 6.3 <sup>bα</sup>	23.7 ± 3.0 <sup>bα</sup>	
J <sub>max</sub> (μmol m <sup>-2</sup> s <sup>-1</sup> )		Winter	Midday	63.4 ± 4.9 <sup>aB</sup>	66.8 ± 2.8 <sup>aB</sup>	—
			Morning	20.0 ± 2.9 <sup>αα</sup>	42.5 ± 4.8 <sup>bα</sup>	46.2 ± 6.7 <sup>bα</sup>
	Summer	Midday	17.1 ± 4.1 <sup>ααA</sup>	34.9 ± 4.3 <sup>bαA</sup>	40.0 ± 9.1 <sup>bα</sup>	
		Evening	16.1 ± 3.3 <sup>αα</sup>	37.8 ± 8.2 <sup>bα</sup>	32.2 ± 4.5 <sup>bα</sup>	
	l <sub>s</sub> (%)	Winter	Midday	26.2 ± 1.5 <sup>bA</sup>	21.9 ± 1.1 <sup>αA</sup>	—
			Morning	58.9 ± 6.3 <sup>bα</sup>	39.9 ± 2.5 <sup>αα</sup>	43.9 ± 1.5 <sup>αα</sup>
Summer		Midday	51.6 ± 3.9 <sup>bαB</sup>	37.5 ± 3.0 <sup>ααB</sup>	42.2 ± 2.1 <sup>αα</sup>	
		Evening	51.5 ± 3.8 <sup>bα</sup>	41.8 ± 1.0 <sup>αα</sup>	37.1 ± 4.3 <sup>αα</sup>	

In all treatments Φ<sub>PSII</sub> and midday F<sub>v</sub>/F<sub>m</sub> values were lower in summer than in winter, whereas NPQ were lower in winter than in summer (Tab. II).

Data derived from the A/C<sub>i</sub> curves performed under midday conditions (Tab. III) showed in winter no difference between R and C in A<sub>max</sub>, V<sub>c,max</sub> or J<sub>max</sub>. There was a decrease from winter to summer, with R showing higher values than C (A<sub>max</sub>: 66.1%, V<sub>c,max</sub>: 57.7%, J<sub>max</sub>: 59.3%, on average). Stomatal limitation (l<sub>s</sub>) was higher in C than in R in both seasons, with l<sub>s</sub> higher in summer than in winter for all treatments (52.1% higher on average). In summer, daily variations were observed for A<sub>max</sub> with the highest values in the morning, whereas no difference was found in V<sub>c,max</sub>, J<sub>max</sub> and l<sub>s</sub>.

### 3.2. Mesophyll conductance

In winter, no difference between treatments was observed in midday g<sub>mes</sub>. In summer, R showed higher daily values than C (Fig. 1). Morning values were 36.1% higher than at midday and in the evening. g<sub>mes</sub> values at midday declined by 97% in controls from winter to summer; whereas in R values declined by 76%. In both seasons, no significant difference in C<sub>C</sub> values between treatments was found (Tab. IV). At midday, the g<sub>mes</sub>/g<sub>s</sub> ratio was higher in winter than in summer in both kinds of plant, whereas no difference was found between treatments in the two seasons (Tab. IV).

### 3.3. Leaf growth parameters

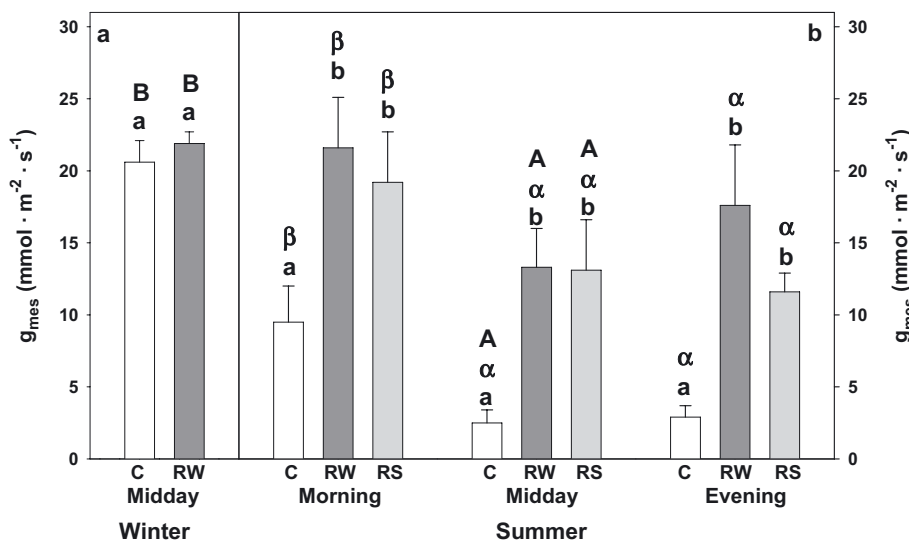
LMA showed no seasonal change. In winter, LMA, *D* and *T* were higher in C (Figs. 2a, 2c, 2d), whereas in summer, R showed lower LMA and *D* but higher *T*. No seasonal difference in density and thickness was found in C. Mean leaf area (LA) was higher in R and decreased from winter to summer (Fig. 2b). LMA and *D* were negatively related to g<sub>mes</sub>; whereas for *T* the relationship was positive (Figs. 3a–3c).

### 3.4. Leaf carbon isotope composition

Isotope discrimination against <sup>13</sup>C (Δ<sup>13</sup>C), calculated from δ<sup>13</sup>C data, was higher in R than in C for both seasons (Tab. V). Results for water-soluble extracts and dry matter showed the same trends. Δ<sup>13</sup>C showed a negative relationship with LMA and a positive relationship with g<sub>mes</sub> for both seasons (Figs. 4a, 4b). Δ<sup>13</sup>C showed a negative relationship with WUE<sub>i</sub> in the winter, whereas in the summer the relationship became positive (Fig. 4c).

## 4. DISCUSSION

In summer, higher temperatures, irradiance and VPD and lower precipitation than in winter lead to increased drought in Mediterranean forests. In fact, g<sub>s</sub>, used as an integrative indicator for the degree of water stress (Galmés et al., 2007), showed



**Figure 1.** Mesophyll conductance to CO<sub>2</sub> ( $g_{mes}$ ), per treatment (control, C; winter resprouts, RW; summer resprouts, RS) and season (winter, summer). In summer, the time of day was included (morning, midday, evening). Values are mean  $\pm$ SE of 10 replicates. Significant differences ( $p \leq 0.05$ ) are indicated by different letters (treatment (a, b, c), season (A, B) and, in summer, time of day ( $\alpha$ ,  $\beta$ ,  $\gamma$ )).

**Table IV.** CO<sub>2</sub> concentration in the chloroplast ( $C_c$ ) and midday mesophyll conductance and stomatal conductance ratio ( $g_{mes}/g_s$ ) at  $C_a = 350 \mu\text{mol mol}^{-1}$  from  $A/C_i$  curves for different treatments (control, C; winter resprouts, RW; summer resprouts, RS) and season (winter, summer). In summer, the time of day was also considered for  $C_c$  values. Each value represents the mean  $\pm$ SE of 10 replicates. Significant differences ( $p \leq 0.05$ ) are indicated by different letters (treatment (a, b, c), season (A, B) and time of the day ( $\alpha$ ,  $\beta$ ,  $\gamma$ )).

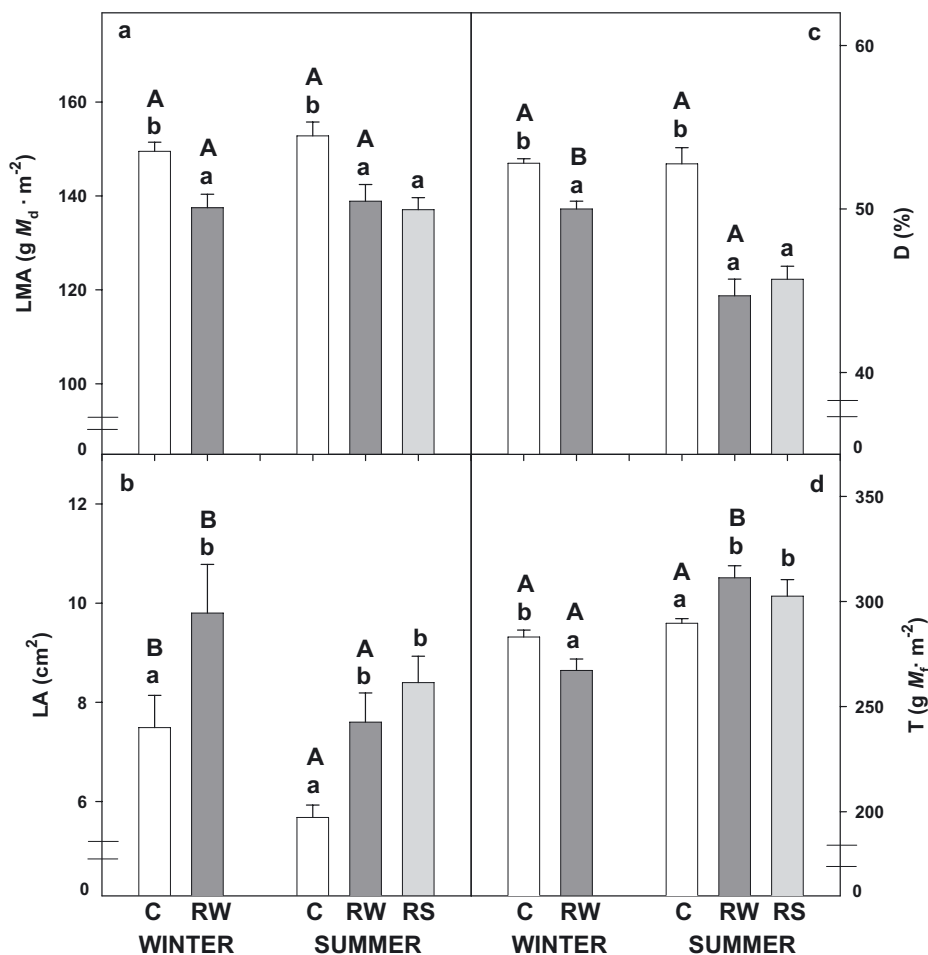
	Season	Time of the day	Treatments		
			C	RW	RS
$C_c$ ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ )	Winter	Midday	75.3 $\pm$ 4.3 <sup>aA</sup>	75.8 $\pm$ 4.0 <sup>aA</sup>	—
		Morning	76.9 $\pm$ 4.0 <sup>aa</sup>	78.5 $\pm$ 2.9 <sup>aa</sup>	84.4 $\pm$ 5.9 <sup>a<math>\beta</math></sup>
	Summer	Midday	66.1 $\pm$ 8.0 <sup>aaA</sup>	90.7 $\pm$ 8.4 <sup>aaA</sup>	96.3 $\pm$ 11.5 <sup>a<math>\beta</math></sup>
		Evening	60.6 $\pm$ 0.8 <sup>aa</sup>	88.2 $\pm$ 9.9 <sup>aa</sup>	67.8 $\pm$ 3.2 <sup>aa</sup>
$g_{mes}/g_s$	Winter	Midday	0.18 $\pm$ 0.03 <sup>aB</sup>	0.17 $\pm$ 0.3 <sup>aB</sup>	—
	Summer	Midday	0.03 $\pm$ 0.01 <sup>aA</sup>	0.04 $\pm$ 0.01 <sup>aA</sup>	0.06 $\pm$ 0.02 <sup>a</sup>

**Table V.** Isotope discrimination against <sup>13</sup>C ( $\Delta^{13}\text{C}$ ), calculated from isotope composition data ( $\delta^{13}\text{C}$ ). Data are shown according to treatments (control, C; winter resprouts, RW; summer resprouts, RS), material analysed (dry matter, DM; water-soluble extract, WSE) and season (winter and summer). Values are mean  $\pm$ SE of 6 replicates. Significant differences across rows or columns ( $p \leq 0.05$ ) are indicated by different letters: treatment (a, b, c), season (A, B) and material analysed ( $\alpha$ ,  $\beta$ ).

	Season	Material analysed	Treatments		
			C	RW	RS
$\Delta^{13}\text{C}$ (‰)	Winter	DM	20.2 $\pm$ 0.25 <sup>aB<math>\alpha</math></sup>	21.5 $\pm$ 0.15 <sup>bB<math>\beta</math></sup>	—
		WSE	19.9 $\pm$ 0.30 <sup>aB<math>\alpha</math></sup>	21.0 $\pm$ 0.14 <sup>bA<math>\alpha</math></sup>	—
	Summer	DM	18.8 $\pm$ 0.18 <sup>aA<math>\alpha</math></sup>	20.2 $\pm$ 0.13 <sup>bA<math>\alpha</math></sup>	20.9 $\pm$ 0.30 <sup>b<math>\alpha</math></sup>
		WSE	18.5 $\pm$ 0.46 <sup>aA<math>\alpha</math></sup>	21.3 $\pm$ 1.42 <sup>bA<math>\alpha</math></sup>	19.8 $\pm$ 0.66 <sup>b<math>\alpha</math></sup>

resprout values corresponding to moderate water stress ( $g_s = 100\text{--}150 \text{ mmol m}^{-2} \text{ s}^{-1}$ ), whereas water stress was severe for undisturbed plants ( $g_s$  below  $50 \text{ mmol m}^{-2} \text{ s}^{-1}$ ). Drought affected numerous measured parameters, declining by 20% in R and by 50% in C:  $A$ ,  $A_{max}$ , diffusive conductance ( $g_s$  and  $g_{mes}$ ),  $V_{c,max}$ ,  $J_{max}$  and  $\Phi_{PSII}$ . Higher values for R in the summer can be explained by the greater nutrient and water availability for

small resprouting shoots than for controls as reflected both in higher  $g_s$  and RWC. Moreover, the larger photosynthetic sink for electrons in R accounts for the lower thermal energy dissipation (estimated by the chlorophyll fluorescence parameter, NPQ) observed in summer, as reported elsewhere (Fleck et al., 1998). In contrast, environmental conditions in the winter did not induce differences in resprouts from undisturbed plants.



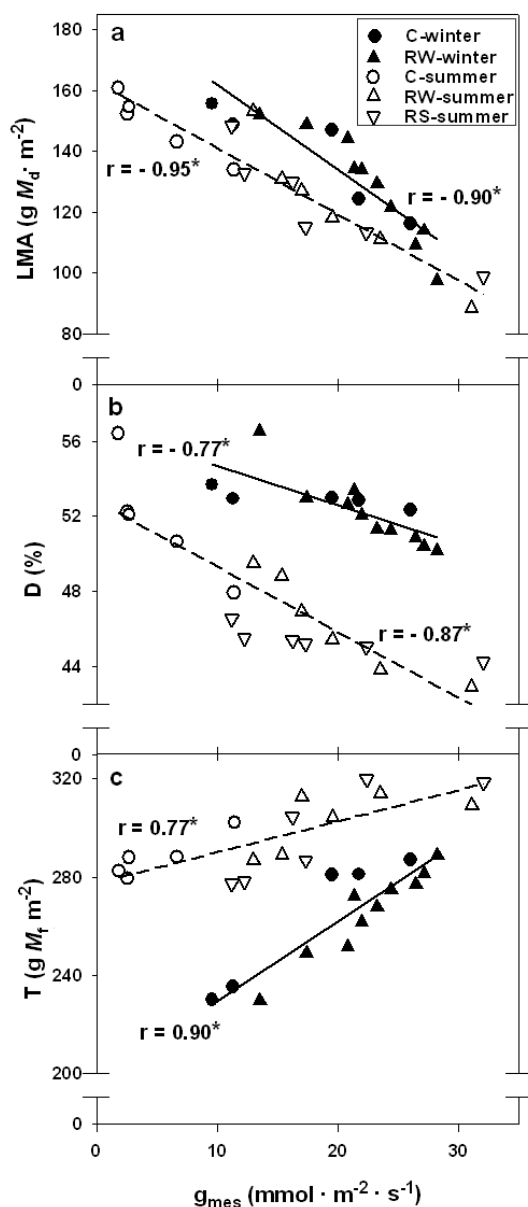
**Figure 2.** Leaf mass per area (LMA), mean area of a leaf (LA), leaf density ( $D$ ) and leaf thickness ( $T$ ) per treatment (control, C; winter resprouts, RW; summer resprouts, RS) in winter and summer. Values are mean  $\pm$ SE of 30 replicates. Significant differences ( $p \leq 0.05$ ) are indicated by different letters (treatment (a, b, c); season (A, B)).

Resprout  $g_{mes}$  was markedly higher (3.75-fold) than in C during summer drought. A decline in  $g_{mes}$  with changes in plant water availability has been observed for other species (Roupsard et al., 1996). The obtained  $g_{mes}$  values were lower than those reported for *Q. ilex* well-watered plants (Loreto et al., 1992). The absolute  $g_{mes}$  values obtained in our study may be under-estimated as some parameters used in the calculations were not measured but assumed from the literature (leaf absorptance, light partition between photosystems I and II) or substituted by approximations (use of dark respiration instead of light respiration). However, our results are in the range obtained by Niinemets et al. (2005) for the same species in a forest study.

The 6-fold decrease in the ratio  $g_{mes}/g_s$  from winter to summer in controls and resprouts suggests a stronger photosynthesis limitation by  $g_{mes}$  in *Q. ilex* than in previously published for other species (Niinemets et al., 2005). During water stress,  $C_i$  may be overestimated because of patchy stomatal closure, and consequently  $g_{mes}$  would be underestimated. However, patchiness was not detected in this study (Materials and Methods, Leaf gas exchange).

The leaf structure of resprouts differed from controls and reflected their higher water availability: in fact, R showed higher mean leaf area and lower LMA, indicative of reduced water stress (Peña-Rojas et al., 2005). These structural characteristics may be primarily responsible for changes in  $g_{mes}$ : the inverse relationship between  $g_{mes}$  and LMA values (Fig. 3a) has been also reported (Niinemets et al., 2006). No seasonal change in LMA was observed in either kind of plants. However, in resprouts, a decrease in density and an increase in thickness were observed from winter to summer. These two components of LMA are not necessarily interdependent, and may be controlled by different environmental variables. However, high  $T$  is commonly associated with lower  $D$  (Mediavilla et al., 2001). In accordance with our results (Figs. 3b, 3c),  $g_{mes}$  reduction has been related to increased  $D$  in peach (Syvertsen et al., 1995), and decreased  $T$  in spinach leaves grown under salt conditions (Delfine et al., 1998).

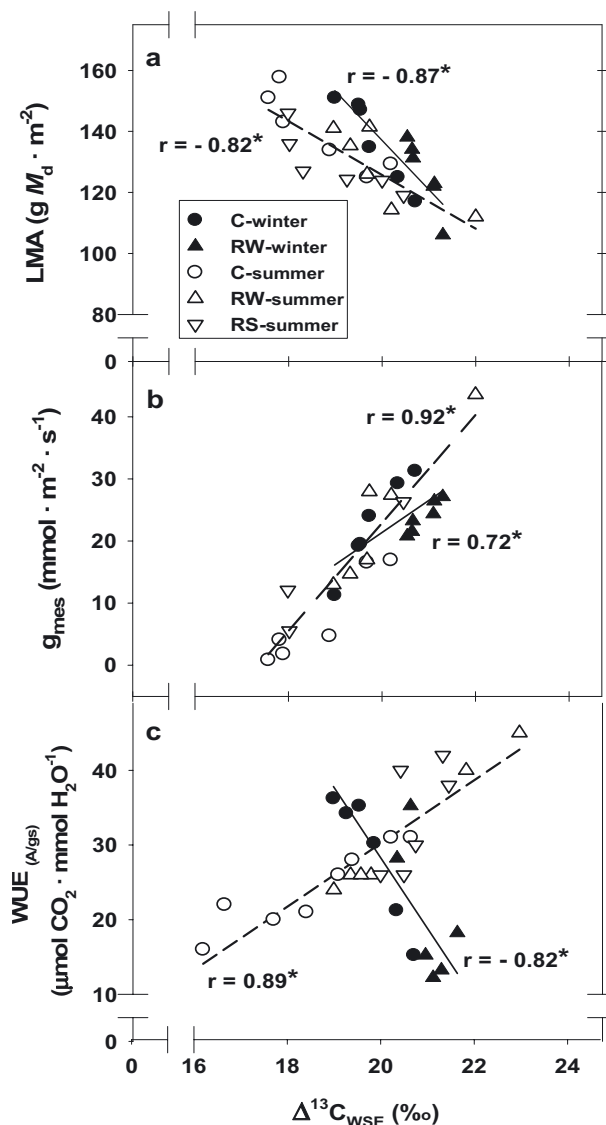
Lower  $D$  and higher  $T$  in resprouts may also account for the observed increased photosynthesis because they correlate with air space fraction in the mesophyll (Niinemets, 1999) resulting in higher  $g_{mes}$ . Moreover,  $T$  is linearly related to the



**Figure 3.** Mesophyll conductance to  $\text{CO}_2$  ( $g_{\text{mes}}$ ) vs. leaf mass per area (LMA) (a), leaf density ( $D$ ) (b) and leaf thickness ( $T$ ) (c) per season (winter, black; summer, white). Symbols represent single measurements of: control (C; ●, ○); winter resprouts, (RW; ▲, △); summer resprouts, (RS; ▽). Asterisks indicate statistically significant correlations ( $* p \leq 0.05$ ).

surface area of cells exposed to intercellular air spaces per unit leaf area (Hanba et al., 2002). As chloroplasts are usually distributed near the cell surface, the  $T$  increase in R accounts for higher photosynthetic protein accumulation per unit leaf area.

However, morphology is not the only factor determining  $g_{\text{mes}}$  since the strong reduction in  $g_{\text{mes}}$  from winter to summer in controls was not paralleled by a change in LMA,  $T$  or  $D$ ; and daily changes in  $g_{\text{mes}}$  in the summer cannot be attributed to changes in leaf morphology, either.  $g_{\text{mes}}$  responds



**Figure 4.** Isotope discrimination against  $^{13}\text{C}$  ( $\Delta^{13}\text{C}$ ) of water-soluble extracts vs. leaf mass per area (LMA) (a),  $g_{\text{mes}}$  (b) and instantaneous water use efficiency ( $\text{WUE}_i = A/g_s$ ) (c) per season (winter, black; summer, white). Symbols represent single measurements of: control (C; ●, ○); winter resprouts, (RW; ▲, △); summer resprouts, (RS; ▽). Asterisks indicate statistically significant correlations ( $* p \leq 0.05$ ).

not only in the long term to environmental stress, but also changes within seconds to minutes even faster than  $g_s$  does (Flexas et al., 2008). Short-term changes in  $g_{\text{mes}}$  have been attributed to carbonic anhydrase (Gillon and Yakir, 2000) and chloroplast aquaporin regulation (Terashima and Ono, 2002; Flexas et al., 2007). Thus, our results are consistent with the idea that  $g_{\text{mes}}$  is grossly determined by leaf structure, but is also the result of physiological control. In our study, the daily variations in  $g_{\text{mes}}$  were of the same magnitude as the seasonal variations, indicating that  $g_{\text{mes}}$  regulation might be as important as the constraints imposed by morphology.



$g_{mes}$  variations in both kinds of plant paralleled changes in  $A$  and  $A_{max}$ , which may indicate a reduction of photosynthesis in response to sustained low chloroplast  $CO_2$  levels (Flexas et al., 2006). However, a limitation of photosynthesis not directly related to  $CO_2$  diffusion is suggested by the analysis of  $A/C_i$  curves. The decreases in  $A$  and  $A_{max}$  in all treatments from winter to summer were paralleled by those of  $V_{c,max}$  and  $J_{max}$ , indicating a non-stomatal limitation of photosynthesis. In  $R$ , this limitation was lower; they showed around 50% higher  $V_{c,max}$  and  $J_{max}$ . These results are compatible with a down-regulation of  $CO_2$  assimilation to adjust mesophyll capacity to the decreased  $CO_2$  supply due to  $g_s$  and  $g_{mes}$  effects (Flexas et al., 2006). This adjustment of the mesophyll capacity would result in maintenance of  $C_c$  as observed (Tab. IV), in the same way that  $C_i$  (Tab. II) tends to remain constant, as reported by Wong et al. (1979).

Morphological and physiological changes during drought can be reflected in  $\Delta^{13}C$  values: we observed a negative relationship between  $\Delta^{13}C$  and LMA values (Fig. 4a), as reported (Fleck et al., 1996). This trend may be a consequence of a  $g_s$  decline, but can also be due to a  $g_{mes}$  decline. A significant contribution of internal resistances to foliar  $\Delta^{13}C$  has been proposed for other species owing to its effect on  $CO_2$  partial pressure at the carboxylation site (Vitousek et al., 1990) and is reflected in Figure 4b. The expected, negative relationship between  $WUE_i$  and  $\Delta^{13}C$  observed in winter values reflects a similar contribution of  $g_s$  and  $g_{mes}$  to  $A$  and  $\Delta^{13}C$ , resulting in  $WUE_i$  increasing as  $g_s$  decreases ( $A$  decreases less than  $g_s$  because of the sustained consumption by RuBisCO) and in  $\Delta^{13}C$  decreasing as  $g_s$  decreases (because of  $C_i$  decline).

The positive relationship observed in summer, (Fig. 4c) can be explained by a dominant role of  $g_{mes}$ , mainly in  $C$ : here,  $A$  declines more than  $g_s$ , especially in some plants, because of the strong reduction in  $g_{mes}$ , resulting in a decrease in  $WUE_i$ , and not in its increase, as expected as  $g_s$  decreases (Tab. II). Meanwhile, the sum of reduced  $g_s$  and  $g_{mes}$ , caused the expected decrease in  $\Delta^{13}C$ , resulting in the observed positive relationship with  $WUE_i$ . Interestingly,  $g_{mes}$  reduction has been proposed as an explanation for the inability of typical gas exchange models to predict  $WUE$  in Mediterranean ecosystems (Reichstein et al., 2002). In fact, Warren and Adams (2006) proposed, from a theoretical point of view, that  $g_{mes}$  may affect the relationship between  $\Delta^{13}C$  and  $WUE$ . Such a disagreement was not found by Roussel et al. (2009) in *Quercus robur*, but Flexas et al. (2008) already found a discrepancy between  $WUE$  and  $\delta^{13}C$  that could be attributed to changes in  $g_{mes}$ , between transgenic tobacco plants, but ours is the first report of a clear mismatch between  $\Delta^{13}C$  and  $WUE$  in forest growing plants that can be attributed to  $g_{mes}$ . The original  $\Delta^{13}C$  model (Farquhar et al., 1982) already included a term for  $g_{mes}$  that is often ignored in typical models, but should be included for prediction of the absolute value of leaf  $\Delta^{13}C$ .

We conclude that  $g_{mes}$  exerts a dominant role in photosynthesis limitation in *Q. ilex*. A regulation of  $g_{mes}$  exists beyond the morphological constraints, and both factors may well be of a similar magnitude. The greater capacity of resprouts to withstand drought that implied lower photosynthetic limitants (both diffusive and non-stomatal) will permit their growth and

recovery after increased fire episodes associated with the climate change.

**Acknowledgements:** This work was supported by funds from the Generalitat de Catalunya (2001SGR00094). We thank L. Cabañeros, J. Vilamú and the Can Coll team (Parc Natural de Collserola) for site facilities; the Servei de Camps Experimentals, UB, for technical assistance; and R. Rycroft and assistants (Servei d'Assessorament Lingüístic, UB) for correcting the English manuscript. K. Peña-Rojas was the recipient of a doctorate grant from AECI and from the Faculty of Forestry Engineering, University of Chile.

## REFERENCES

- Balaguer L., Afif D., Dizengremel P., and Dreyer E., 1996. Specificity factor of ribulose biphosphate carboxylase/oxygenase of *Quercus robur*. *Plant Physiol. Biochem.* 34: 879–883.
- Brooks A. and Farquhar G.D., 1985. Effect of temperature on the  $CO_2/O_2$  specificity of ribulose-1,5-bisphosphate carboxylase oxygenase and the rate of respiration in the light – estimates from gas-exchange measurements on spinach. *Planta* 165: 397–406.
- Christensen J.H., Hewitson B., Busuioc A., Chen A., Gao X., Held I., Jones R., Kolli R.K., Kwon W.T., Laprise R., Rueda V.M., Mearns L., Menéndez C.G., Räisänen J., Rinke A., Sarr A., and Whetton P., 2007. Regional Climate Projections. In: Solomon S., Qin D., Manning M., Chen Z., Marquis M., Averyt K.B., Tignor M., and Miller H.L. (Eds.), *Climate Change 2007: The Physical Science Basis. Contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change*, Cambridge University Press, Cambridge, United Kingdom and New York, pp. 847–940.
- Delfine S., Alvino A., Zacchini M., and Loreto F., 1998. Consequences of salt stress on conductance to  $CO_2$  diffusion, Rubisco characteristics and anatomy of spinach leaves. *Aust. J. Plant Physiol.* 25: 395–402.
- Epron D., Godard D., Cornic G., and Genty B., 1995. Limitation of net  $CO_2$  assimilation rate by internal resistances to  $CO_2$  transfer in the leaves of two tree species (*Fagus sylvatica* L. and *Castanea sativa* Mill.). *Plant Cell Environ.* 18: 43–51.
- Ethier G.J. and Livingston N.J., 2004. On the need to incorporate sensitivity to  $CO_2$  transfer conductance into Farquhar-von Caemmerer-Berry leaf photosynthesis model. *Plant Cell Environ.* 27: 137–153.
- Farquhar G.D., O'Leary M.H., and Berry J.A., 1982. On the relationship between carbon isotope discrimination and the intercellular carbon dioxide concentration in leaves. *Aust. J. Plant Physiol.* 9: 121–137.
- Farquhar G.D., Ehleringer J.R., and Hubick K.T., 1989. Carbon isotope discrimination and photosynthesis. *Ann. Rev. Plant Physiol. Plant Mol. Biol.* 40: 503–537.
- Fleck I., Grau D., Sanjosé M., and Vidal D., 1996. Carbon isotope discrimination in *Quercus ilex* resprouts after fire and tree-fell. *Oecologia* 105: 286–292.
- Fleck I., Hogan K.P., Llorens L., Abadía A., and Aranda X., 1998. Photosynthesis and photoprotection in *Quercus ilex* resprouts after fire. *Tree Physiol.* 18: 607–614.
- Flexas J., Ribas-Carbó M., Bota J., Galmés J., Henkle M., Martínez-Cañellas S., and Medrano H., 2006. Decreased Rubisco activity during water stress is not induced by decreased relative water content but related to conditions of low stomatal conductance and chloroplast  $CO_2$  concentration. *New Phytol.* 172: 73–82.
- Flexas J., Diaz-Espejo A., Galmés J., Kaldenhoff R., Medrano H., and Ribas-Carbó M., 2007. Rapid variations of mesophyll conductance in response to changes in  $CO_2$  concentration around leaves. *Plant Cell Environ.* 30: 1284–1298.

- Flexas J., Ribas-Carbó M., Diaz-Espejo A., Galmés J., and Medrano H., 2008. Mesophyll conductance to CO<sub>2</sub>: current knowledge and future prospects. *Plant Cell Environ.* 31: 602–621.
- Galmés J., Medrano H., and Flexas J., 2007. Photosynthetic limitations in response to water stress and recovery in Mediterranean plants with different growth forms. *New Phytol.* 175: 81–93.
- Gillon J.S. and Yakir D., 2000. Internal conductance to CO<sub>2</sub> diffusion and (CO<sub>2</sub>)-O<sup>18</sup> discrimination in C<sub>3</sub> leaves. *Plant Physiol.* 123: 201–213.
- Hanba Y.T., Kogami H., and Terashima I., 2002. The effects of growth irradiance on leaf anatomy and photosynthesis in *Acer* species differing in light demand. *Plant Cell Environ.* 25: 1021–1030.
- Harley P.C., Loreto F., Di Marco G., and Sharkey T.D., 1992. Theoretical considerations when estimating the mesophyll conductance to CO<sub>2</sub> flux by the analysis of the response of photosynthesis to CO<sub>2</sub>. *Plant Physiol.* 98: 1429–1436.
- Laing W.A., Ögren W.L., and Hegeman R.H., 1974. Regulation of soybean net photosynthetic CO<sub>2</sub> fixation by the interaction of CO<sub>2</sub>, O<sub>2</sub> and ribulose 1,5 diphosphate carboxylase. *Plant Physiol.* 54: 678–685.
- Lawlor D.W. and Tezara W., 2009. Causes of decreased photosynthetic rate and metabolic capacity in water-deficient leaf cells: a critical evaluation of mechanisms and integration processes. *Ann. Bot.* 103: 561–579.
- Loreto F., Harley P.C., Di Marco G., and Sharkey T.D., 1992. Estimation of mesophyll conductance to CO<sub>2</sub> flux by three different methods. *Plant Physiol.* 98: 1437–1443.
- Mediavilla S., Escudero A., and Heilmeyer H., 2001. Internal leaf anatomy and photosynthetic resource-use efficiency interspecific and intraspecific comparisons. *Tree Physiol.* 21: 251–259.
- Niinemets Ü., 1999. Research review: Components of leaf dry mass per area – Thickness and density – later leaf photosynthetic capacity in reverse directions in woody plants. *New Phytol.* 144: 35–47.
- Niinemets Ü., Cescatti A., Rodeghiero M., and Tosens T., 2005. Leaf internal diffusion conductance limits photosynthesis more strongly in older leaves of Mediterranean evergreen broad-leaved species. *Plant Cell Environ.* 28: 1552–1566.
- Niinemets Ü., Cescatti A., Rodeghiero M., and Tosens T., 2006. Complex adjustments of photosynthetic potentials and internal diffusion conductance to current and previous light availabilities and leaf age in Mediterranean evergreen species in *Quercus ilex*. *Plant Cell Environ.* 29: 1559–1578.
- Peña-Rojas K., Aranda X., and Fleck I., 2004. Stomatal limitation to CO<sub>2</sub> assimilation and down-regulation of photosynthesis in *Quercus ilex* L. resprouts under slowly-imposed drought. *Tree Physiol.* 24: 813–822.
- Peña-Rojas K., Aranda X., Joffre R., and Fleck I., 2005. Leaf morphology, photochemistry and water status changes in resprouting *Quercus ilex* during drought. *Funct Plant Biol.* 32: 117–130.
- Reichstein M., Tenhunen J.D., Rouspard O., Ourcival J.M., Rambal S., Miglietta F., Peressotti A., Pecchiari M., Tirone G., and Valentini R., 2002. Severe drought effects on ecosystem CO<sub>2</sub> and H<sub>2</sub>O fluxes at three Mediterranean evergreen sites: revision of current hypotheses? *Global Change Biol.* 8: 999–1017.
- Rouspard O., Gross P., and Dreyer E., 1996. Limitation of photosynthetic activity by CO<sub>2</sub> availability in the chloroplasts of oak leaves from different species and during drought. *Ann. Sci. For.* 53: 243–254.
- Roussel M., Dreyer E., Montpied P., Le-Provost G., Guehl J.-M., and Brendel O., 2009. The diversity of <sup>13</sup>C isotope discrimination in a *Quercus robur* full-sib family is associated with differences in intrinsic water use efficiency, transpiration efficiency, and stomatal conductance. *J. Exp. Bot.* 60: 2419–2431.
- Syvertsen J.P., Lloyd J., McConchie C., Kriedemann P.E., and Farquhar G.D., 1995. On the relationship between leaf anatomy and CO<sub>2</sub> diffusion through the mesophyll of hypostomatous leaves. *Plant Cell Environ.* 18: 149–157.
- Terashima I., Miyazawa S.-I., and Hanba Y.T., 2001. Why are sun leaves thicker than shade leaves? Consideration based on analysis of CO<sub>2</sub> diffusion in the leaf. *J. Plant Res.* 114: 93–105.
- Terashima I. and Ono K., 2002. Effects of HgCl<sub>2</sub> on CO<sub>2</sub> dependence of leaf photosynthesis: evidence indicating involvement of aquaporins in CO<sub>2</sub> diffusion across the plasma membrane. *Plant Cell Physiol.* 43: 70–78.
- Vitousek P.M., Field C.B., and Matason P.A., 1990. Variation in foliar δ<sup>13</sup>C in Hawaiian *Metrosideros polymorpha*: a case of internal resistance? *Oecologia* 84: 362–370.
- Warren C.R. and Adams M.A., 2006. Internal conductance does not scale with photosynthetic capacity: implications for carbon isotope discrimination and the economics of water and nitrogen use in photosynthesis. *Plant Cell Environ.* 19: 192–201.
- Wong S.C., Cowan I.R., and Farquhar G.D., 1979. Stomatal conductance correlates with photosynthetic capacity. *Nature* 282: 424–426.